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Total synthesis of mycestericin A and its 14-epimer

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1. Introduction

Mycestericin A (1) is a member of mycestericins produced by Mycelia sterilia and has been reported to have a potent immuno-suppressive activity.^{[1](#page-13-0)} A structural elucidation study revealed that the structure of mycestericin A is similar to that of myriocin, a wellknown immunosuppressant and serine palmitoyltransferase (SPT) inhibitor, possessing an intriguing α -substituted α -amino acid structure,^{[2](#page-13-0)} although mycestericin A has another E-olefin between C-12 and C-13 and one distal (R) -allylic alcohol function at C-14 ([Fig. 1](#page-1-0)).^{1a} The stereochemistry at C-14 was determined using the benzoate CD chirality method of an N-acetyl-14-O-benzoyl derivative of mycestericin A γ -lactone, which was prepared from natural **1.**^{[1a](#page-13-0)} Mycestericin A and its congeners have been reported to have comparable IC_{50} values to that of myriocin in the nanomolar range in the mouse allogeneic mixed lymphocyte reaction.^{[1](#page-13-0)} Due to their potent biological properties such as immunosuppressive, antifungal, and SPT inhibitory activities, 3 naturally occurring molecules of this type, including myriocin,² sphingofungins,⁴ sulfa-misterins,^{[5](#page-13-0)} and mycestericins,¹ as well as their derivatives are expected to be promising lead compounds for novel therapeutic agents based on their ability to modulate sphingolipid biosynthesis. 6 These interesting biological findings as well as architecturally novel structures have stimulated a number of synthetic efforts, including total syntheses and synthetic approaches to

ABSTRACT

The total synthesis of mycestericin A (1) and its 14-epimer 34 is described herein. The Overman rearrangement of an allylic trichloroacetimidate derived from L-tartrate generated a tetra-substituted carbon with nitrogen and subsequent stereoselective transformations afforded the highly functionalized left-half segment, vinyl iodide. Cross-coupling of the vinyl iodide with a chiral organometallic species synthesized from p-tartrate under the Negishi or Suzuki–Miyaura coupling conditions, followed by deprotection, completed the total synthesis of 1. The 14-epimer of mycestericin A was also synthesized, and a comparison of $\alpha|_D$ values of peracetyl γ -lactone derivatives of mycestericin A and its 14-epimer as well as degradation studies of 1 and 34 fully confirmed the proposed absolute structure of mycestericin A. - 2009 Elsevier Ltd. All rights reserved.

> generate natural products possessing a-substituted a-amino acid structures.[7–11](#page-13-0)

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To date, however, there has been no reported study of the synthesis of mycestericin A, which possesses the most complicated structure in the mycestericin family. In this paper, we present the first report of the total synthesis of mycestericin A and its 14-epimer from tartrates, and the confirmation of the proposed absolute structure of the natural product.¹²

2. Results and discussion

2.1. Retrosynthesis

Our previous success with the total synthesis of myriocin, [9e,10h](#page-13-0) sphingofungin E,^{10h} and lactacystin,¹³ from aldohexoses suggested that the Overman rearrangement^{14,15} on chiral scaffolds would effectively generate the tetra-substituted carbon with nitrogen. This idea involves disconnection of the carbon framework in 1 into the highly functionalized left-half segment, vinyl iodide 2 or 3, and the hydrophobic right-half segment, organometallic species 4, possessing an (E,R)-allylic alcohol function ([Fig. 2](#page-1-0)). The well-established Pd-catalyzed coupling reaction of 2 or 3 with 4 was expected to stereoselectively construct the carbon backbone with two E-olefins in 1. We planned to prepare the left-half segment 2 or 3 from homoallyl alcohol 6, which was thought to arise by the stereoselective allylation of aldehyde 7. The Overman rearrangement of chiral allylic trichloroacetimidate 8 would install a tetra-substituted carbon with nitrogen in 7, generating a precursor of α -substituted α amino acid. For the preparation of imidate 8, L-tartrate was chosen

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Figure 1. Structures of mycestericin A and related natural products.

as a homochiral starting material. On the other hand, the counterpart 4, which would be generated from alkyl iodide 5, was planned to be synthesized from D -tartrate by the E-selective carbon-elongation reaction.

2.2. Preparation of the highly functionalized left half, vinyl iodide 2

The synthesis of 2 commenced with the known acetonide 9^{16} 9^{16} 9^{16} prepared from dimethyl L -tartrate in a three-step reactions^{[11e](#page-13-0)} with a 31% overall yield (Scheme 1). After protection of the primary hydroxy group in 9, the O-benzyl group in 10 was removed to give 11 with a 94% yield from 9. PCC oxidation of 11, followed by the Wittig reaction generated 12 as the single isomer (93% for two steps). The E-geometry of 12 was confirmed by the observed NOE between a vinyl proton and H-4. The reduction of 12 with DIBAL generated allyl alcohol 13 with a yield of 97%. The treatment of 13 with trichloroacetonitrile and DBU produced trichloroacetimidate 8, which, without purification, was heated in xylene in the presence of $K_2CO_3^{17}$ $K_2CO_3^{17}$ $K_2CO_3^{17}$ in a sealed tube at 140 °C for 48 h. Due to Overman rearrangement, this reaction provided 14[18](#page-13-0) and its epimer, epi-14, with isolated yields of 62% and 33%, respectively. Although the observed stereoselectivity in the Overman rearrangement of 8 was moderate ($14:epi-14=1.9:1$), the relatively short-step synthesis of 14 from 9 with acceptable overall yields led us to employ this approach for the total synthesis. At this stage, the structure of 14 could not be fully determined but was later confirmed by X-ray analysis of the more advanced compound 6 (vide infra).

Scheme 1. Bn=-CH₂Ph, PCC=pyridinium chlorochromate, MS4A=molecular sieves 4 Å, DIBAL= $[(CH_3)_2CHCH_2]_2$ AlH.

The transformation of rearranged product 14 to generate 2 was next examined [\(Scheme 2\)](#page-2-0). Ozonolysis of 14, followed by further oxidation and esterification produced 15 with a 97% yield. Removal of the O-silyl protecting group resulted in 16 (100% yield). Pfitzner– Moffatt oxidation of 16 gave aldehyde 7, which, without purification, was reacted with allyl tributyltin in the presence of $MgBr₂$ in CH_2Cl_2 ^{[19](#page-13-0)} to stereoselectively generate an allylated product, homoallyl alcohol 6, as the sole product in 75% yield from 16.

The chelation control (between an aldehyde-carbonyl and an α -ether oxygen) was expected to favor the formation of 6, the structure of which has been unambiguously assigned by a single crystal X-ray analysis.²⁰ The hydroxy group in 6 was protected as Figure 2. Retrosynthetic route to mycestericin A. MOM=-CH₂OMe, TBDPS=-SiPh₂(t-Bu). a methoxymethyl ether to give 17 (80% yield). Ozonolysis of 17,

Scheme 3. 9-BBN=9-borabicyclo[3.3.1]nonane, Ms= $-SO₂Me$.

Scheme 2. DCC=N,N'-dicyclohexylcarbodiimide, TFA=CF₃CO₂H.

followed by the Takai reaction^{[21](#page-13-0)} with CHI₃ in the presence of CrCl₂ in THF/1,4-dioxane provided (E) -vinyl iodide (2) with a 60% yield as a geometrically pure compound after chromatographic purification.

2.3. Synthesis of the hydrophobic right-half segment

The counterpart for the coupling reaction, a precursor of organometallic species, iodide $5a$, was synthesized from D -tartrate (Scheme 3). The known di-O-tosylate²² **18**, prepared from diisopropyl p-tartrate with an 86% overall yield, was treated with n pentylmagnesium bromide in the presence of CuBr to afford 19 in 53% yield. After deprotection of the acetonide group, the product was converted into epoxide 20 (93% yield), whose hydroxy group was protected as an MOM ether to give 21a (96% yield).

Oxidative cleavage of the epoxide group in 21a with $HIO₄²³$ $HIO₄²³$ $HIO₄²³$ generated the corresponding aldehyde, which was then reacted with the stabilized Wittig reagent to afford an inseparable mixture of E-olefin and its Z-isomer ($E:Z=6.5:1$). DIBAL reduction of the mixture followed by chromatographic separation gave geometrically pure E-allyl alcohol 22a in 55% isolated yield from 21a. The primary hydroxy group in 22a was transformed into O-mesylate, which was then reacted with allylmagnesium chloride^{[24](#page-13-0)} to provide 23a in 81% yield. Hydroboration of 23a with excess 9-BBN (8 equiv to 23a) followed by oxidation provided primary alcohol 24a, whose hydroxy function was replaced with iodide, to give $5a¹⁸$ $5a¹⁸$ $5a¹⁸$ a precursor for the coupling reaction, with a 75% yield.

2.4. Attempted synthesis of 1 via the Negishi coupling reaction

Having established the procedure for the preparation of both counterparts for the coupling reaction, we subsequently explored the conditions of the Negishi cross-coupling reaction, 25 25 25 which has been utilized for the total synthesis of myriocin and sphingofungin F by Ham et al. 9f,10i 9f,10i 9f,10i The reaction of iodide **5a** with *t-*BuLi at -78 $^{\circ}$ C followed by treatment with $ZnCl₂$ generated an alkyl zinc species. This was then reacted with vinyl iodide 2 in the presence of $Pd(PPh_3)_4$ to generate the coupling product, fully protected mycestericin A (25), in 86% yield (Scheme 4). The treatment of 25 with aqueous HCl at 60° C removed all the protecting groups, however, the concomitant elimination of a methoxymethyloxy or hydroxy function at C-14 (mycestericin A numbering) also occurred and, after acetylation, γ -lactone-diene 27^{[26](#page-13-0)} was obtained. On the

Scheme 4.

other hand, the reaction of 25 with aqueous acetic acid followed by acetylation afforded a γ -lactone, which is expected to possess the structure of 28b (49% yield). The $^1\mathrm{H}$ NMR data of the synthetic γ -lactone were very similar to those reported for the lactone 28b derived from natural mycestericin A^{1a} A^{1a} A^{1a} and the FABMS data also supported its structure. However, in the ¹³C NMR of the synthetic γ -lactone, in addition to a set of four signals (δ 123.2, 128.6, 134.0, and 135.0 ppm) due to olefinic carbons whose chemical shifts are in good agreement with those of the authentic 28b, an extra four signals (δ 123.1, 128.1, 134.4, and 134.6 ppm) of olefinic carbons, whose intensities were almost same as those of the former four signals, were observed. These results revealed that the synthetic γ -lactone is an inseparable mixture of diastereomers. The most plausible products of the acid hydrolysis of 25, followed by acetylation, are 28a,b and 29a,b, which would be formed via the allyl cation intermediate 26. The attempted deprotection of 25 with TMSBr, which was reported to be effective for the clean deprotection of an allylic MOM ethers,²⁷ also resulted in the formation of a mixture of 28a,b and 29a,b. These results clearly showed that the allylic alcohol moiety in 25 is unexpectedly labile under the acidic conditions used, prompting us to devise an improved route to 1.

2.5. The alternative approach to 1: completion of the total synthesis

To avoid the formation of the allyl cation 26, a coupling reaction of substrates possessing protecting groups that could be removed under basic conditions was next explored (Scheme 5). For this purpose, compound 6 was converted into γ -lactone 30 (98% yield) by treatment with aqueous acetic acid at 70° C followed by acetylation. Ozonolysis of 30 and subsequent Takai reaction generated a new left-half segment, vinyl iodide 3, as a single E-isomer after chromatographic purification with a yield of 73%. On the other hand, for the preparation of the right-half segment, the alcohol function in epoxide 20 was protected as a 2-(trimethylsilyl)ethoxymethyl (SEM) group to give 21b (100% yield). Then, the same reaction sequence as employed for the conversion of 21a to 5a was applied to 21b to provide $5b^{18}$ $5b^{18}$ $5b^{18}$ with a 28% overall yield from 21b. Another right-half segment 23c, possessing a TBDPS protecting group, was also synthesized from 20 (56% overall for six steps) following the same reaction sequence. Although epoxide 20 could also be transformed into its OTBS and OTES derivatives, it was found that the periodate oxidation of the epoxides possessing OTBS and OTES groups resulted in the decomposition of the substrates.

The Negishi coupling of 5b-derived alkyl zinc with γ -lactone 3, followed by acetylation, successfully provided 31 (44% yield; Scheme 6). Compound 31 was treated with anhydrous Bu₄NF and MS4A in N,N-dimethyl propylene urea (DMPU)^{[28](#page-13-0)} at 80 °C and then acetylated. Under these reaction conditions, the SEM protecting group was successfully removed without formation of an allyl cation to generate tetraacetyl mycestericin A γ -lactone (28b) as a single isomer (38% yield). The spectral data (1 H and 13 C NMR) as well as the α _D value of the synthetic 28b were identical with those reported for γ -lactone obtained from natural mycestericin A.^{[1a](#page-13-0)} Finally, alkaline hydrolysis of 28b followed by ion-exchange resin (IRC-76) treatment furnished mycestericin A (1) in 93% yield. The [α]_D value { α]²¹ -8.6 (c 0.45, MeOH); lit.^{[1a](#page-13-0)} [α]²⁵ -8.5 (c 0.50, MeOH)} and spectroscopic data showed good concordance with those reported for the natural product.^{[1a](#page-13-0)} Thus, the first total synthesis of mycestericin (1) has been achieved.

2.6. The second-generation total synthesis via the Suzuki– Miyaura coupling

As described in the previous section, mycestericin A (1) was successfully synthesized using Negishi coupling as the key reaction. However, the yield of the coupling reaction (44%) was not satisfactory and, furthermore, the final deprotection also resulted in

Scheme 6. DMPU=N,N-dimethyl propylene urea.

a poor yield (38%). The low yield of the Negishi coupling may be due to the presence of base-sensitive functionalities (γ -lactone and acetyl groups) in the left-half segment (3), which might have undergone attack by an excess of organozinc or organolithium reagents. The harsh basic reaction conditions for the deprotection of the OSEM group would have induced the decomposition of the substrate and/or product.²⁹ To improve these crucial steps, the Suzuki–Miyaura method 30 30 30 of coupling γ -lactone **3** with an organoborane derived from the alkene 23c possessing OTBDPS group was next investigated. The B-alkyl Suzuki–Miyaura coupling procedure has been successfully employed in the total synthesis of sphingofungins E and F by Trost,^{[10d](#page-13-0)} and sphingofungin E by Nakamura and Shiozaki[.10g](#page-13-0)

Hydroboration of 23c with excess 9-BBN followed by H_2O quench produced the corresponding organoborane, which was then reacted with 3 in the presence of $PdCl₂ \cdot dppf$, $Ph₃As$, and $K₂CO₃$ in DMF (Scheme 7). To our delight, under these reaction conditions, coupling product 32 was obtained with a yield of 87%. The use of Ph₃As was found to be essential for a higher yield; the similar reaction in the absence of Ph₃As gave 32 in 58% yield.³¹ Although the treatment of 32 with Bu₄NF in THF at room temperature resulted in recovery of the starting material, the reaction performed at 65 °C generated 28b in 63% yield after acetylation. By use of the Suzuki–Miyaura coupling reaction, the overall yield of mycestericin A was much improved (4.3% yield from 9 using the Negishi coupling approach compared to 14.0% yield from 9 with the Suzuki–Miyaura approach).

2.7. Total synthesis of 14-epi-mycestricin A (34): confirmation of the stereochemistry at C-14 of the natural product

As described in the [Introduction,](#page-0-0) the C-14 stereochemistry of mycestericin A had been determined by way of the benzoate CD chirality method for the N-acetyl-14-O-benzoyl derivative of mycestericin A γ -lactone.^{1a} To confirm the stereochemistry at C-14 by the synthetic method, synthesis of 14-epimer of mycestericin A was next investigated. For this purpose, the enantiomer of diene **23c** (ent-**23c**) was synthesized starting from dimethyl L -tartrate by a similar reaction sequence to that employed for the conversion of diisopropyl D-tartrate to 23c (Scheme 8).^{[32](#page-13-0)} The B-alkyl Suzuki-Miyaura coupling of an organoborane generated from ent-23c with vinyl iodide 3 afforded the coupling product 33 in 77% yield. Deprotection of the OTBDPS group and subsequent acetylation provided 14-epi-tetraacetyl mycestericin A γ -lactone (28a) in 26% yield. Alkaline hydrolysis of 28a, followed by ion-exchange resin (IRC-76) treatment, resulted in an 81% yield of 14-epi-mycestericin A (34).

The $\rm ^1H$ and $\rm ^{13}C$ NMR spectra of 28a and 34 were quite similar to those of 28b and 1, respectively, and we were unable to detect any notable differences between the NMR spectra of 28a and 28b and those between 34 and 1. Fortunately, it was found that the $[\alpha]_D$ values of these compounds displayed some differences as shown in Table 1. The relatively large differences in $\alpha|_D$ values between 28b and 28a (Table 1, entries 1–3) allowed us to clearly distinguish these two compounds, assigning the stereochemistry at C-14 in natural mycestericin A as R.

Table 1 $[\alpha]_D$ values of compounds 28b, 28a, 1, and 34

Entry	Compound Source		$\lceil \alpha \rceil_D$	Measurement conditions
	28 b	From natural product $+58.4^a$ 25 °C, c 0.5, CHCl ₃		
2	28 _b	Synthetic		+58.2 21 °C, c 0.41, CHCl ₃
3	28a	Synthetic		+30.0 21 °C, c 0.22, CHCl ₃
4	1	Natural product		-8.5^a 25 °C, c 0.5, MeOH
5	1	Synthetic		-8.6 21 °C, c 0.45, MeOH
6	34	Synthetic	-5.0	20 °C, c 0.15, MeOH

^a Ref. [1a.](#page-13-0)

2.8. Degradation studies of mycestericin A (1) and its 14-epimer (34)

It is a possible that the allylic hydroxy functions at C-14 in compounds 1 and 34 might undergo partial epimerization during the synthetic or purification process via an allyl cation intermediate and it is now understood that such epimerization cannot be detected by 1 H nor 13 C NMR analyses. To confirm the stereochemical purities at C-14 in final compounds 1 and 34, studies of their degradation were conducted [\(Scheme 9](#page-5-0)). In the preparation of reference compounds (36 and ent-36) for HPLC analyses, compound 22b was converted into diacetate 35 (77% yield). Ozonolysis

of 35 (reductive workup with NaBH4), followed by the removal of O-acetyl groups and subsequent O-benzoylation, produced authentic 36 in 49% yield. A similar reaction sequence was applied to ent-22b to give ent-36. The chiral HPLC analyses (DAICEL CHIR-ALCEL OJ-H, 4.6 mm ID \times 250 mm, *i*-PrOH/hexane=1:300, flow rate=0.8 mL/min) of 36 (retention time: 21.9 min) and ent-36 (retention time: 19.0 min) revealed that the optical purities of both compounds were >99% ee. Next, the degradation reactions of synthetic mycestericin A (1) and its 14-epimer (34) were performed. Treatment of 1 with Ac_2O /pyridine gave tetraacetyl mycestericin A γ -lactone (28b), whose ozonolysis (reductive workup with NaBH4) followed by basic methanolysis and subsequent benzoylation provided a dibenzoate in 50% overall yield after chromatographic separation. The same reaction sequence was applied to 14-epi-mycestericin $A(34)$ to afford a dibenzoate via 28a in 38% overall yield. The $^1\mathrm{H}$ NMR spectra of these dibenzoates were totally identical to those of 36. The chiral HPLC analyses of degradation products clearly showed that the dibenzoate derived from mycestericin A (1) is 36 with $>99\%$ ee, whereas the dibenzoate obtained from 14-epi-mycestericin A (34) is ent-36 with >99% ee. From these experiments it was confirmed that no epimerization at C-14 had occurred during the synthetic and purification processes for the preparation of 1 and 34, and our final products (1 and 34) were both diastereomerically homogeneous.

dimethyl $n - C_nH₁₃$ \longleftarrow ent-22b \longrightarrow ent-35 \rightarrow $-$ BzO D-tartrate **OBz** $ent-36$

(>99%ee; retention time of chiral HPLC, 19.0 min)

3. Conclusion

In summary, the first total synthesis of mycestericin $A(1)$ from dimethyl L-tartrate has been achieved via an efficient route (24 steps and 4.3% overall yield). The synthesis of 14-epi-mycestericin A (34), comparison of the $[\alpha]_D$ values of their γ -lactone derivatives (28b and 28a), and the degradation studies of 1 and 34 fully confirmed the proposed absolute structure of the natural product, including the stereochemistry at the distal C-14 hydroxy group. This work has also provided a new synthetic pathway for the synthesis of highly oxygenated α -substituted α -amino acid derivatives with potent biological activities utilizing readily available tartrates.

4. Experimental

4.1. General

Melting points were determined on a Mitamura-Riken micro hot stage and were not corrected. Optical rotations were recorded using a sodium lamp (589 nm) with a JASCO DIP-370 instrument with 1 dm tube and values of $[\alpha]_D$ are recorded in units of 10^{-1} deg cm² g⁻¹. Infrared (IR) spectra were measured with a JASCO FT/IR-200 spectrometer. ¹H NMR spectra were recorded at 300 MHz on a JEOL Lambda 300 or on a Varian MVX-300 spectrometers for solutions in CDCl₃, unless otherwise noted. Chemical shifts are reported as δ values in parts per million. Abbreviations used are: b (broad peak), s (singlet), d (doublet), t (triplet), q (quartet) and m (complex multiplet). 13C NMR spectra were recorded at 75 MHz on a JEOL Lambda 300 spectrometer for solutions in CDCl₃, unless otherwise noted. Chemical shifts are reported as δ values in parts per million. Mass spectra are measured by a JEOL GC Mate spectrometer with EI (70 eV) or FAB mode. Organic extracts were dried over solid anhydrous $Na₂SO₄$ and concentrated below 40 °C under reduced pressure. Column chromatography was carried out with silica gel (Merck Kieselgel 60 F₂₅₄; 230-400 mesh) for purification. Preparative TLC (PLC) was performed with Merck PLC plate (Kieselgel 60 F_{254} , 0.5 mm thickness).

4.2. Synthesis of the left-half segment

4.2.1. {[(4S,5S)-5-(Benzyloxy)-2,2-dimethyl-1,3-dioxan-4-yl]methoxy}- (tert-butyl)-diphenylsilane (10). To a solution of [(4S,5S)-5-(benzyloxy)-2,2-dimethyl-1,3-dioxan-4-yl]methanol^{16,11e} (9) (1.34 g, 5.31 mmol) in DMF (18 mL) at room temperature were added imidazole (545 mg, 8.01 mmol) and TBDPSCl (1.90 mL, 7.31 mmol). After being stirred at room temperature for 20 h, the reaction mixture was diluted with brine, and products were extracted with EtOAc. The organic layer was washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (100 g silica gel, 1:20–1:10 EtOAc/hexane as an eluent) to afford **10** (2.61 g, 100%) as a colorless syrup: $\lbrack \alpha \rbrack_0^{25}$ +15.3 (c 0.89, CHCl₃); IR (neat) 3450, 2990, 2940, 2875, 1455, 1385, 1200, 1120, 1080, 1055, 1025 cm⁻¹; ¹H NMR δ 1.06 (9H, s), 1.39, 1.40 (each 3H, 2s), 3.43 (1H, ddd, J = 1.4, 2.0, 3.9 Hz), 3.71 (1H, ddd, J = 3.9, 9.0, 9.0 Hz), 3.89 (1H, dd, J=2.0, 12.9 Hz), 3.94-4.02 (3H, m), 4.56 and 4.72 (each 1H, 2d, J=11.9 Hz), 7.25–7.46 (11H, m), 7.63–7.68 (4H, m); ¹³C NMR δ 19.1, 19.4, 27.1, 29.2, 62.2, 63.0, 69.7, 71.6, 72.3, 98.7, 127.7, 127.9, 128.0, 128.5, 129.8, 129.9, 133.7, 133.8, 135.8, 135.8, 138.6; HRMS (FAB) m/z calcd for C₃₀H₃₈O₄SiNa (M+Na)⁺ 513.2437, found: m/z 513.2452.

4.2.2. (4S,5S)-4-[(tert-Butyldiphenylsilyloxy)methyl]-2,2-dimethyl-1,3-dioxan-5-ol (11). To a solution of naphthalene (1.02 g, 7.96 mmol) in THF at -10 °C was added Li (37 mg, 5.3 mmol), and the mixture was stirred at -10 °C for 30 min. To this mixture -10 °C under Ar was added a solution of 10 (261 mg, 0.532 mmol) in THF (5.6 mL). After being stirred at -10 °C for 3.5 h, the reaction mixture was quenched by addition of EtOH (2 mL) and then diluted with EtOAc. The organic layer was washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (50 g silica gel, 1:40–1:3 EtOAc/hexane as an eluent) to afford 11 (200 mg, 94%) as a colorless syrup: $\lbrack \alpha \rbrack^{22}_{\text{D}} + 15.6$ (c 1.52, CHCl₃); IR ν_{max} (neat) 3480, 2995, 2960, 2930, 2860, 1470, 1425, 1365, 1275, 1230, 1200, 1130, 1110, 1065 cm⁻¹; ¹H NMR δ 1.06 (9H, s), 1.40 and 1.43 (each 3H, 2s), 2.76 (1H, d, $J=10.2$ Hz), 3.63 (1H, dddd, J = 1.5, 1.5, 2.7, 10.2 Hz), 3.68 (1H, dd, J = 5.1, 9.8 Hz), 3.85 (1H, dd, J = 2.7, 12.2 Hz), 3.87 (1H, dd, J = 7.1, 9.8 Hz), 3.95 (1H, ddd, J = 1.5, 5.1, 7.1 Hz), 4.05 (1H, dd, J=1.5, 12.2 Hz), 7.34-7.46 (6H, m), 7.65-7.74 (4H, m); ¹³C NMR δ 18.6, 19.4, 27.0, 29.6, 63.7, 66.2, 72.4, 99.1, 127.9, 129.9, 133.4, 133.7, 135.8, 135.9; HRMS (FAB) m/z calcd for $C_{23}H_{33}O_4Si$, $(M+H)^+$ 401.2148, found: 401.2145. Anal. Calcd for C23H32O4Si: C, 68.83; H, 8.05. Found: C, 68.83; H, 7.98.

4.2.3. (2E)-Ethyl 2-{(4R)-4-[(tert-butyldiphenylsilyloxy)methyl]-2,2 dimethyl-1,3-dioxan-5-ylidene}acetate (12). To the mixture of 11 (1.04 g, 2.60 mmol) and MS 4 Å (800 mg) in CH₂Cl₂ (40 mL) was added a suspension of PCC (1.96 g, 9.10 mmol), NaOAc (1.49 g, 18.2 mmol), and MS 4 Å (800 mg) in CH_2Cl_2 (40 mL), and the mixture was stirred at room temperature for 2 h. The reaction mixture was partially concentrated and then diluted with $Et₂O$. The insoluble material was removed by filtration through a pad of Celite. The filtrate was concentrated to give a crude ketone (929 mg) as a yellow syrup, which was used for next reaction without further purification. To a solution of the crude ketone (929 mg) in toluene (27 mL) was added $Ph_3P=CHCO_2Et$ (2.70 g, 7.80 mmol), and the mixture was stirred at $100\degree C$ for 12 h. The mixture was concentrated to give a residue, which was purified by column chromatography (50 g silica gel, 1:20 EtOAc/toluene as an eluent) to afford **12** (1.14 g, 93% for two steps) as a colorless syrup: $[\alpha]_D^{22}$ – 52.4 (*c* 0.70, CHCl₃); IR v_{max} (neat) 2990, 2935, 2860, 1715, 1415, 1370, 1210, 1150, 1115 cm⁻¹; ¹H NMR δ 1.06 (9H, s), 1.27 (3H, t, J=7.3 Hz), 1.35 and 1.36 (each 3H, 2s), 3.83 (1H, dd, J=5.9, 10.5 Hz), 3.90 (1H, dd, J=5.6, 10.5 Hz), 4.15 (2H, q, J=7.3 Hz), 4.41 (1H, ddd, J=2.0, 5.6, 5.9 Hz), 4.61 $(1H, ddd, J=2.0, 3.9, 17.8 Hz)$, 5.02 $(1H, dd, J=2.0, 17.8 Hz)$, 5.78 $(1H,$ dd, 1H, J=2.0, 3.9 Hz) 7.34–7.46 (6H, m), 7.64–7.71 (4H, m); ¹³C NMR d 14.4, 19.4, 24.4, 25.3, 27.0, 60.3, 61.9, 65.0, 71.7, 100.3, 112.6, 127.9, 127.9, 129.9, 133.5, 135.8, 135.8, 158.9, 166.1; HRMS (FAB) m/z calcd for C₂₇H₃₇O₅Si, $(M+H)^+$ 469.2410, found: 469.2407. Anal. Calcd for $C_{27}H_{36}O_5Si$: C, 69.20; H, 7.74. Found: C, 69.19; H, 7.54.

4.2.4. (2E)-2-{(4R)-4-[(tert-Butyldiphenylsilyloxy)methyl]-2,2-dimethyl-1,3-dioxan-5-ylidene}ethanol (13). To a solution of 12 $(4.09 \text{ g}, 8.73 \text{ mmol})$ in toluene (80 mL) at $-78 \text{ }^{\circ}\text{C}$ was added DIBAL (1.01 M solution in toluene, 21.6 mL, 21.8 mmol) dropwise under Ar. After being stirred at -78 °C for 1 h, to the reaction mixture at -78 °C was added acetone (10 mL), and the mixture was further stirred at $0 °C$ for 10 min. The reaction mixture was diluted with EtOAc, and washed successively with 1 M aqueous HCl solution, saturated aqueous NaHCO₃ solution and brine, and dried. Removal of the solvent gave a residue, which was purified by column chromatography (150 g silica gel, 1:3 EtOAc/hexane as an eluent) to afford $\bf 13$ (3.62 g 97%) as a colorless syrup: $[\alpha]_D^{21}$ –40.4 (c 1.75, CHCl₃); IR v_{max} (neat) 3400, 2930, 2860, 1475, 1425, 1380, 1370, 1225, 1135, 1115, 1080, 1010 cm⁻¹; ¹H NMR δ 1.07 (9H, s), 1.36 and 1.41 (each 3H, 2s), 3.82 (1H, dd, J=6.1, 10.7 Hz), 3.92 (1H, dd, J=4.9, 10.7 Hz), 4.00-4.17 (2H, m), 4.27 and 4.36 (each 1H, 2d, $J=14.4$ Hz), 4.41 (1H, ddd, J=1.7, 4.9, 6.1 Hz), 5.49 (1H, dt, J=1.7 and 6.1 Hz), 7.34–7.46 (6H, m), 7.66–7.74 (4H, m); ¹³C NMR δ 19.4, 23.5, 26.6, 27.0, 58.3, 59.4, 65.7, 72.1, 99.9, 121.4, 127.8, 127.8, 129.8, 129.9, 133.7, 133.8, 135.8, 135.9, 138.2; HRMS (FAB) m/z calcd for C₂₅H₃₄O₄SiNa, $(M+Na)^+$ 449.2124, found: 449.2126.

4.2.5. N-{(4R,5S)-4-[(tert-Butyldiphenylsilyloxy)methyl]-2,2-dimethyl-5-vinyl-1,3-dioxan-5-yl}-2,2,2-trichloroacetamide (14) and its $(4R,5R)$ -isomer (epi-14). To a solution of 13 (741 mg, 1.74 mmol) in CH_2Cl_2 (20 mL) at 0 °C were added DBU (26.0 μ L, 0.17 mmol) and trichloroacetonitrile (0.440 mL, 4.34 mmol), and the mixture was stirred at $0°C$ for 10 min. The reaction mixture was concentrated to give a residue, which was passed through a short column of silica gel (8 g, 1:5 EtOAc/hexane containing 1% Et3N as an eluent) to afford crude imidate 8 (961 mg) as a pale yellow syrup. A mixture of crude **8** (961 mg) and K_2CO_3 (48 mg) in o-xylene (96 mL) was heated in a sealed tube at 140° C for 48 h. The reaction mixture was concentrated to give a residue, which was purified by column chromatography (50 g silica gel, 1:20 EtOAc/hexane) to afford first, **14** (615 mg, 62% from 12) as a colorless syrup. Further elution gave

epi-14 (327 mg, 33% from 12) as a crystalline residue. Data for 14: $[\alpha]_D^{21}$ +12.6 (c 1.20, CHCl₃); IR ν_{max} (neat) 3400, 2930, 2860, 1720, 1515, 1505, 1425, 1380, 1200, 1140, 1115 cm⁻¹; ¹H NMR δ 1.08 (9H, s), 1.47 and 1.48 (each 3H, 2s), 3.80 (1H, d, J=11.9 Hz), 3.80-3.91 (2H, m), 3.94 (1H, dd, J=2.2, 10.2 Hz), 4.31 (1H, d, J=11.9 Hz), 5.16 (1H, d, J=17.5 Hz), 5.28 (1H, d, J=11.2 Hz), 5.78 (1H, dd, J=11.2, 17.5 Hz), 7.32–7.48 (6H, m), 7.64–7.74 (5H, m); ¹³C NMR δ 18.5, 19.4, 27.0, 29.2, 58.3, 63.0, 64.2, 75.8, 92.9, 99.5, 116.9, 127.7, 127.7, 129.8, 133.1, 133.3, 135.7, 135.8, 161.2; HRMS (FAB) m/z calcd for C₂₇H₃₅Cl₃NO₄Si, $(M+H)^+$ 570.1401, found: 570.1398. Anal. Calcd for C₂₇H₃₄NO₄Cl₃Si: C, 56.79; H, 6.00; N, 2.45. Found: C, 56.61; H, 6.29; N, 2.45. Data for *epi*-**14**: mp 77.6–79.0 °C; $[\alpha]_D^{25}$ +1.8 (*c* 0.47, CHCl₃); IR (neat) 3340, 2915, 1860, 1730, 1510, 1430, 1240, 1200, 1100, 1055 cm $^{-1}$; ¹H NMR δ 1.11 (9H, s), 1.38 and 1.51 (each 3H, 2s), 3.66 (1H, dd, J=3.9, 10.2 Hz), 3.88 (1H, dd, J=8.3, 10.2 Hz), 4.12 (1H, d, J=11.7 Hz), 4.19 $(1H, dd, J=3.9, 8.3 Hz), 4.61 (1H, d, J=11.7 Hz), 5.36 (1H, d,$ J=17.6 Hz), 5.41 (1H, d, J=11.0 Hz), 6.25 (1H, dd, J=11.0, 17.6 Hz), 7.34–7.48 (6H, m), 7.58–7.66 (4H, m), 7.70 (1H, br s); ¹³C NMR δ 18.9, 19.5, 27.3, 28.6, 58.5, 65.3, 66.1, 72.8, 99.3, 115.9, 127.9, 130.1, 132.7, 132.7, 132.8, 135.5, 161.3; HRMS (FAB) m/z calcd for C₂₇H₃₅Cl₃NO₄Si, $(M+H)^+$ 570.1401, found: 570.1392. Anal. Calcd for C₂₇H₃₄NO₄Cl₃Si: C, 56.79; H, 6.00; N, 2.45. Found: C, 56.99; H, 6.12; N, 2.30.

Starting from diisopropyl D-tartrate, an enantiomer of 14 (ent-14) was synthesized by the similar procedures as described for the preparation of 14 from dimethyl L-tartrate. Optical purities of 14 and ent-14 were determined to be >99% ee by chiral HPLC analyses [DAICEL CHIRALCEL OD-H, 4.6 mm ID × 250 mm, i-PrOH/ hexane=1:15, flow rate=0.8 mL/min, UV (254 nm) detection]: retention time for 14, 6.0 min; for ent-14, 7.2 min.

4.2.6. Methyl (4R,5S)-4-[(tert-butyldiphenylsilyloxy)methyl]-2,2-dimethyl-5-(2,2,2-trichloroacetamido)-1,3-dioxane-5-carboxylate (15). Ozone was introduced into a solution $14(375 \text{ mg}, 0.657 \text{ mmol})$ in EtOH (6.6 mL) at -78 °C for 25 min. After confirming the complete consumption of the starting material (TLC analysis), excess ozone was removed with a stream of Ar gas. To the reaction mixture was added Me₂S (0.48 mL, 6.6 mmol) at -78 °C and the mixture was stirred at room temperature for 5 h. The resulting mixture was diluted with EtOAc and washed with brine, and dried. Removal of the solvent gave a crude aldehyde (400 mg) as a pale yellow syrup. To a solution of the crude aldehyde (400 mg) in t-BuOH (4.4 mL) and H₂O (4.4 mL) were added NaH₂PO₄ \cdot 2H₂O (206 mg, 1.32 mmol), $HOSO₂NH₂$ (192 mg, 1.98 mmol), and NaClO₂ (239 mg, 2.64 mmol). After being stirred at room temperature for 15 h, the reaction mixture was diluted with 20 wt% aqueous $Na₂S₂O₃$ solution, and products were extracted with $CHCl₃$ five times. The combined organic layers were dried and concentrated to give a crude carboxylic acid (540 mg) as a white solid. To a solution of the crude carboxylic acid in MeOH (1.7 mL) and benzene (6.6 mL) at 0° C was added (trimethylsilyl)diazomethane $(2.0 M$ solution in Et₂O, 0.43 mL, 0.86 mmol), and the mixture was stirred at room temperature for 1.5 h. Removal of the solvent gave a residue, which was purified by column chromatography (15 g silica gel,1:5 EtOAc/hexane) to afford **15** (382 mg, 97%) as a colorless syrup: $[\alpha]_D^{20}$ +16.7 (*c*, CHCl₃); IR ν_{max} (neat) 3410, 2955, 2930, 1740, 1715, 1505, 1430, 1385, 1255, 1200, 1115, 1060 cm⁻¹; ¹H NMR δ 1.08 (9H, s), 1.47 and 1.49 (each 3H, 2s), 3.72 (3H, s), 3.79 (1H, dd, J=4.4, 11.5 Hz), 3.96 (1H, dd, J=3.9, 11.5 Hz), 4.20 (1H, dd, J=3.9, 4.4 Hz), 4.27 and 4.34 (each 1H, 2d, J=12.4 Hz), 7.34–7.45 (6H, m), 7.65–7.71 (4H, m), 7.86 (br s, 1H, NH); ¹³C NMR d 18.9, 19.6, 27.2, 28.9, 53.1, 60.9, 62.9, 63.7, 73.0, 92.3, 99.9, 127.9, 127.9,130.1,133.2,133.2,135.9,161.8,168.6; HRMS (FAB) m/z calcd for $C_{27}H_{35}Cl_{3}NO_{6}Si$, $(M+H)^{+}$ 602.1300, found: 602.1285.

4.2.7. Methyl (4R,5S)-2,2-dimethyl-4-(hydroxymethyl)-5-(2,2,2-trichloroacetamido)-1,3-dioxane-5-carboxylate (16). To a solution of 15 (382 mg, 0.633 mmol) in THF (6.3 mL) was added Bu₄NF (1.0 M

solution in THF, 1.10 mL, 1.10 mmol) at -15 °C. After being stirred at -15 °C for 1 h, the reaction mixture was diluted with EtOAc, and washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (6 g silica gel, 1:2– 1:1 EtOAc/hexane as an eluent) to afford 16 (231 mg, 100%) as a crystalline residue: mp 57.3–58.9 °C; [α] $_{\rm D}^{28}$ +38.1 (c 1.20, CHCl $_{\rm 3}$); IR v_{max} (neat) 3570, 3480, 3300, 1735, 1715, 1655, 1520, 1390, 1260, 1205, 1125, 1085 cm⁻¹; ¹H NMR δ 1.46 and 1.53 (each 3H, 2s), 2.38 (1H, br s), 3.76–3.84 (1H, m), 3.77 (3H, s), 3.90 (1H, dd, \approx 1=3.4, 12.4 Hz), 4.15 (1H, dd, J=3.2, 3.4 Hz), 4.34 (2H, s), 8.60 (br s, 1H, NH); ¹³C NMR δ 19.2, 28.8, 53.3, 61.4, 62.3, 62.8, 71.4, 92.4, 100.0, 162.1, 168.9; HRMS (FAB) m/z calcd for C₁₁H₁₇Cl₃NO₆, $(M+H)^+$ 364.0122, found: 364.0123.

4.2.8. Methyl (4R,5S)-2,2-dimethyl-4-[(R)-1-hydroxybut-3-enyl]-5-(2,2, 2-trichloroacetamido)-1,3-dioxane-5-carboxylate (6). To a solution of 16 (249 mg, 0.683 mmol) in benzene (6.8 mL) at room temperature were added pyridine (0.055 mL, 0.68 mmol), TFA (0.026 mL, 0.34 mmol), DMSO (3.4 mL), and DCC (561 mg, 2.72 mmol). After being stirred at room temperature for 5 h, the reaction mixture was diluted with Et₂O, and washed successively with water and brine, and dried. The organic layer was concentrated to give crude aldehyde 7 (589 mg), which was used for the next reaction without further purification. To a solution of $MgBr₂$ (0.068 mL, 1.37 mmol) in $CH₂Cl₂$ (7.0 mL) at room temperature was added a solution of crude aldehyde **7** (589 mg) in CH_2Cl_2 (6.6 mL) via a cannula under Ar. The mixture was stirred at -78 °C for 10 min. To this mixture at -78 °C was added allyl tributyltin (0.32 mL, 1.02 mmol). After stirring at -78 °C for 15 min, the cooling bath was removed, and the resulting mixture was stirred at ambient temperature for 25 h. The reaction mixture was diluted with 1 M aqueous HCl solution at 0° C, and products were extracted with EtOAc. The organic layer was washed successively with 1 M aqueous HCl solution, saturated aqueous $NAHCO₃$ solution and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (25 g silica gel, 1:30 EtOAc/toluene as an eluent) to give 6 (207 mg, 75%) as a crystalline residue: mp 98.2–99.2 °C (from EtOAc); [α] $^{22}_{\rm D}$ +56.1 (c 1.04, CHCl₃); IR $\nu_{\rm max}$ (neat) 3580, 3305, 1750, 1710, 1540, 1385, 1245, 1200 cm $^{-1}$; 1 H NMR δ 1.44 and 1.47 (each 3H, 2s), 2.26 (1H, ddd, J=7.1, 7.1, 13.9 Hz), 2.35–2.45 $(2H, m)$, 3.76 $(3H, s)$, 3.89 $(1H, ddd, J=1.0, 7.1, 16.1 Hz)$, 3.94 $(1H, d, d)$ $J=1.0$ Hz), 4.36 (1H, d, $J=12.4$ Hz), 4.45 (1H, d, $J=12.4$ Hz), 5.11 (1H, d, J=16.8 Hz), 5.13 (1H, d, J=8.5 Hz), 5.68 (1H, m), 9.22 (1H, br s); ¹³C NMR d 19.4, 28.1, 39.2, 53.1, 62.3, 62.4, 69.9, 70.8, 92.3, 99.9, 119.2, 132.8, 162.0, 169.1; HRMS (FAB) m/z calcd for C₁₄H₂₁Cl₃NO₆, (M+H)⁺ 404.0434, found: 404.0428. Anal. Calcd for $C_{14}H_{20}NO_6Cl_3$: C, 41.55; H, 4.98; N, 3.46. Found: C, 41.66; H, 4.99; N, 3.48.

4.2.9. Methyl (4R,5S)-2,2-dimethyl-4-[(R)-1-(methoxymethoxy)but-3-enyl]-5-(2,2,2-trichloroacetamido)-1,3-dioxane-5-carboxylate (17). To a solution of 6 (25 mg, 0.062 mmol) in $(CH_2Cl)_2$ (1 mL) at room temperature were added i -Pr₂NEt (0.12 mL, 0.68 mmol), MOMCl (0.028 mL, 0.37 mmol), and tetrabutylammonium chloride (2.5 mg, 0.007 mmol). After stirring at room temperature for 17 h, the reaction mixture was diluted with EtOAc, and washed successively with saturated aqueous $NH₄Cl$ solution, saturated aqueous $NaHCO₃$ solution and brine, and dried. Removal of the solvent gave a residue, which was purified by column chromatography (1 g silica gel, 1:7 EtOAc/hexane as an eluent) to afford 17 (22 mg, 79%) as a colorless syrup: [α] $^{30}_{\rm D}$ +88.9 (c 0.77, CHCl3); IR $\nu_{\rm max}$ (neat) 3365, 2995, 2955, 1730, 1715, 1515, 1505, 1385, 1260, 1200 cm⁻¹; ¹H NMR δ 1.46 and 1.49 (each 3H, 2s), 2.44–2.54 (2H, m), 3.43 (3H, s), 3.72–3.80 (1H, m), 3.78 $(3H, s)$, 4.08 (1H, d, J=1.0 Hz), 4.25 and 4.54 (each 1H, 2d, J=12.2 Hz), 4.78 and 4.82 (each 1H, 2d, J=7.1 Hz), 5.11 (1H, d, J=16.1 Hz), 5.12 (1H, d, $J=10.2$ Hz), 5.58–5.64 (1H, m), 8.73 (1H, br s); ¹³C NMR δ 18.8, 29.1, 35.4, 53.1, 56.7, 62.4, 62.5, 71.1, 76.7, 92.7, 96.5,100.2,119.1,133.1,161.3, 168.8; HRMS (FAB) m/z calcd for C₁₆H₂₅Cl₃NO₇, (M+H)⁺ 448.0697, found: 448.0686. Anal. Calcd for C₁₆H₂₄NO₇Cl₃: C, 42.83; H, 5.39; N, 3.12. Found: C, 43.00; H, 5.51; N, 3.02.

4.2.10. Methyl (4R,5S)-2,2-dimethyl-4-[(R,E)-4-iodo-1-(methoxymethoxy)but-3-enyl]-5-(2,2,2-trichloroacetamido)-1,3-dioxane-5-carboxylate (2). Ozone was introduced into a solution of 17 (32 mg, 0.071 mmol) in EtOH (2 mL) at -78 °C for 1.5 min. After purging of excess ozone with a stream of Ar gas, to the reaction mixture at -78 °C was added Me₂S (0.052 mL, 0.71 mmol), and the resulting mixture was stirred at room temperature for 5 h. The mixture was diluted with EtOAc and washed with brine, and dried. Removal of the solvent gave a crude aldehyde (35 mg) as a pale yellow syrup. To a suspension of $CrCl₂$ (349 mg, 2.84 mmol) in THF (3.5 mL) and 1,4dioxane (3.5 mL) at 0 \degree C under Ar was added a solution of the crude aldehyde (35 mg) and CHI₃ (335 mg, 0.85 mmol) in THF (3.5 mL) via a cannula. After being stirred at room temperature for 20 h, the reaction was quenched by the addition of $H₂O$ (1 mL). The resulting mixture was diluted with $Et₂O$ and washed successively with 30 wt % aqueous $Na₂S₂O₃$ solution and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (1.5 g, silica gel, 1:5 EtOAc/hexane as an eluent) to afford 2 (28 mg, 60% from **17**) as a colorless syrup: [α] $_D^{30}$ +61.0 (c 0.82, CHCl3); IR v_{max} (neat) 3370, 2915, 2850, 1730, 1715, 1515, 1380, 1260, 1150 cm⁻¹; ¹H NMR δ 1.46 and 1.49 (each 3H, 2s), 2.42-2.60 (2H, m), 3.42 (3H, s), 3.75–3.83 (1H, m), 3.81 (3H, s), 4.01 (1H, d, J=1.0 Hz), 4.26 and 4.54 (each 1H, 2d, $J=12.2$ Hz), 4.77 and 4.80 (each 1H, 2d, $J=7.1$ Hz), 6.15 (1H, d, $J=14.4$ Hz), 6.40 (1H, ddd, $J=6.8$, 8.3, 14.4 Hz), 8.64 (1H, br s); ¹³C NMR δ 18.5, 29.0, 38.0, 53.1, 56.6, 62.1, 62.2, 71.5, 75.8, 78.4, 96.8, 100.1, 100.5, 140.6, 161.1, 168.5; HRMS (FAB) m/z calcd for C₁₆H₂₄Cl₃INO₇, $(M+H)^+$ 573.9664, found: 573.9650.

4.2.11. (2R,3R,4S)-4-Acetamido-4-(acetoxymethyl)-2-allyl-5-oxotetrahydrofuran-3-yl acetate (30). A solution of 6 (185 mg, 0.457 mmol) in AcOH (16 mL) and H_2O (4 mL) was stirred at 80 $\,^{\circ}$ C for 11 h. The reaction mixture was concentrated to give a residue, which was dissolved in pyridine (6 mL) and Ac_2O (5 mL). After stirring at room temperature for 9 h, the reaction mixture was concentrated to give a residue, which was purified by column chromatography (4 g silica gel, 1:5–2:3 EtOAc/hexane as an eluent) to give 30 (141 mg, 98%) as a colorless syrup: $[\alpha]_D^{22}$ +71.4 (c 0.63, CHCl₃). IR $\nu_{\rm max}$ (neat) 3350, 3270, 3050, 2930, 1780, 1755, 1750, 1670, 1540, 1375, 1230, 1185, 1050, 1030 cm⁻¹; ¹H NMR δ 2.03, 2.05 and 2.10 (each 3H, 3s), 2.37–2.56 (2H, m), 4.50 (2H, s, 2H), 4.77 (1H, td, J=4.9, 8.3 Hz), 5.13-5.21 (2H, m), 5.75–5.89 (2H, m), 6.07 (1H, br s); 13 C NMR δ 20.3, 20.5, 22.7, 33.2, 62.6, 62.8, 72.0, 80.9,118.8,132.0,168.9,169.4,170.1,172.2; HRMS (FAB) m/z calcd for C₁₄H₁₉NO₇, $(M+H)^+$ 314.1240, found: 314.1242.

4.2.12. (2R,3R,4S)-4-Acetamido-4-(acetoxymethyl)-2-[(E)-3-iodoallyl]-5-oxotetrahydrofuran-3-yl acetate (3). Ozone was introduced into a solution of 30 (113 mg, 0.361 mmol) in EtOH (10 mL) at -78 °C for 10 min. After confirming the complete consumption of the starting material (TLC analysis), excess ozone was removed with a stream of Ar gas. To the reaction mixture was added Me₂S (0.30 mL, 4.1 mmol) at -78 °C and the mixture was stirred at $0 °C$ for 6 h. The reaction mixture was concentrated to give a residue, which was diluted with EtOAc. The organic layer was washed with brine, and dried. Removal of the solvent left a crude aldehyde (115 mg). To a suspension of $CrCl₂$ (442 mg, 3.60 mmol) in dioxane (6.0 mL) under Ar at $0 °C$ was added a THF solution (3.0 mL) of the crude aldehyde (115 mg) and CHI₃ (433 mg) 1.10 mmol) via a cannula. After stirring at room temperature for 1.5 h, the reaction was quenched by addition of 20 wt % aqueous Na₂S₂O₃ solution (20 mL), and then diluted with EtOAc. The organic layer was washed successively with 20 wt % aqueous $Na₂S₂O₃$ solution, $H₂O$ and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (3 g silica gel,

1:2–2:1 EtOAc/hexane as an eluent) to give first, the Z-isomer of 3 (20 mg, 13%) as a colorless syrup. Further elution afforded 3 (115 mg, 73%) as a crystalline residue. Data for $3:$ mp 58.0–58.9 °C; $[\alpha]_D^{24}$ +83.5 (c 0.93, CHCl₃); IR $\nu_{\rm max}$ (KBr disk) 3350, 3050, 1780, 1755, 1750, 1680, 1540, 1375, 1230, 1185, 1050, 1030 cm⁻¹; ¹H NMR d 2.03, 2.06 and 2.12 (each 3H, 3s), 2.38–2.56 (2H, m), 4.49 (2H, s), 4.77 (1H, ddd, J=4.9, 8.6, 8.6 Hz), 5.75 (1H, d, J=4.9 Hz), 5.99 (1H, br s), 6.27 (1H, ddd, J=1.2, 1.2, 14.4 Hz), 6.55 (ddd, 1H, J=7.1, 7.1, 14.4 Hz); ¹³C NMR δ 20.8, 21.1, 23.1, 35.9, 62.6, 63.8, 72.3, 79.5, 79.8, 140.1, 169.7, 170.2, 170.5, 170.7; HRMS (FAB) m/z calcd for $C_{14}H_{18}NO_7$ INa, $(M+Na)^+$ 462.0026, found: 462.0031. Data for Zisomer of **3**: $[\alpha]_D^{19}$ +67.1 (c 0.24, CHCl₃); IR ν_{max} (neat) 3350, 3050, 1780, 1755, 1750, 1680, 1540, 1375, 1230, 1185, 1050 cm⁻¹; ¹H NMR δ 2.03, 2.09 and 2.10 (each 3H, 3s), 2.51–2.68 (2H, m), 4.50 (2H, s), 4.84 (1H, ddd, J=4.6, 5.2, 5.2 Hz), 5.79 (1H, d, J=4.6 Hz), 6.06 (1H, br s), 6.31 (1H, ddd, J=6.8, 6.8, 7.2 Hz), 6.46 (1H, ddd, J=1.5, 1.5, 7.2 Hz); 13 C NMR (75 MHz, CDCl₃) δ 20.4, 20.6, 22.6, 34.5, 62.4, 62.9, 71.8, 86.3, 134.8, 169.0, 169.5, 170.3, 172.0; HRMS (FAB) m/z calcd for $C_{14}H_{19}NO_{7}I$, $(M+H)^{+}$ 440.0206, found: 440.0214.

4.3. Synthesis of the right-half segment

4.3.1. [(4R,5R)-2,2-Dimethyl-5-hexyl-1,3-dioxolan-4-yl]methanol 4 methylbenzenesulfonate (19). To a suspension of CuBr (1.30 g, 9.03 mmol) in THF (30 mL) at 0 \degree C under Ar was added *n*-pentylmagnesium bromide (1.5 M solution in Et₂O, 34.4 mL, 51.6 mmol), and the mixture was cooled to -30 °C. To this suspension was added a THF solution (22 mL) of [(4R,5R)-2,2-dimethyl-5-hexyl-1,3 dioxolan-4-yl]-4,5-dimethanol 4,5-bis(4-methylbenzenesulfo-nate)^{[22](#page-13-0)} (18) (6.09 g, 12.9 mmol). After being stirred for 19 h at -30 °C, the reaction mixture was diluted with 1 M aqueous HCl solution (30 mL) at -30 °C, and products were extracted with EtOAc. The organic layer was washed successively with 1 M aqueous HCl solution, saturated aqueous NaHCO₃ solution and brine, and dried. Removal of the solvent gave a residue, which was purified by column chromatography (150 g silica gel, 1:10 EtOAc/hexane as an eluent) to afford **19** (2.55 g, 53%) as a colorless syrup: [α] $_{\text{D}}^{\text{21}}$ +19.9 (c 1.44, CHCl₃); IR ν_{max} (neat) 2935, 2855, 1370, 1190, 1180, 1100, 980 cm⁻¹; ¹H NMR δ 0.88 (3H, t, J=6.9 Hz), 1.24-1.58 (10H, m), 1.29 and 1.35 (each 3H, 2s), 2.45 (3H, s), 3.74–3.82 (2H, m), 4.05 (1H, dd, $J=4.1$, 10.5 Hz), 4.11 (1H, dd, $J=3.7$, 10.5 Hz), 7.34 and 7.79 (each 2H, 2d, J=8.0 Hz); ¹³C NMR δ 14.2, 21.8, 22.7, 26.0, 26.9, 27.5, 29.4, 31.8, 33.2, 69.4, 78.0, 78.4, 109.5, 128.2, 130.0, 132.9, 145.2; HRMS (EI) m/z calcd for C₁₉H₃₀O₅S, (M)⁺ 370.1814, found: 370.1820.

4.3.2. (R)-1-[(R)-Oxiran-2-yl]heptan-1-ol (20). A solution of 19 (6.50 g, 17.5 mmol) in AcOH (80 mL) and H_2O (20 mL) was heated at 70 \degree C for 13 h. The reaction mixture was concentrated to give a crude diol (6.12 g) as a crystalline residue, which was dissolved in $CH₂Cl₂$ (88 mL). To this solution at room temperature was added DBU (7.35 mL, 49.1 mmol), and the resulting mixture was stirred at room temperature for 2 h. After addition of saturated aqueous NH4Cl solution (40 mL), the reaction mixture was diluted with EtOAc, and washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (65 g silica gel, 1:15–1:4 EtOAc/hexane as an eluent) to give 20 (2.58 g, 93%) as a colorless syrup: [α] $_{\rm D}^{23}$ –3.6 (c 2.51, CHCl $_{\rm 3}$); IR $\nu_{\rm max}$ (neat) 3440, 3955, 2930, 2860, 1730, 1460, 1255, 1085, 980 cm $^{-1};\,{}^{1}\text{H}\,{}{\rm NMR}$ δ 0.88 (3H, t, J=7.1 Hz), 1.20–1.67 (10H, m), 1.82 (1H, br s), 2.72 (1H, dd, J=2.7, 4.9 Hz), 2.82 (1H, dd, J=4.1, 4.9 Hz), 2.98 (1H, ddd, J=2.7, 4.1, 5.0 Hz), 3.44 (1H, dt, J=5.0, 5.9 Hz); ¹³C NMR (75 MHz, CDCl₃) d 14.2, 22.7, 25.4, 29.4, 31.8, 34.4, 45.3, 55.7, 71.9; HRMS (FAB) m/z calcd for C₉H₁₉O₂, $(M+H)^+$ 159.1385, found: 159.1394.

4.3.3. (R)-2- $[(R)-1-(Methoxymethoxy)heptyl]oxirane (21a)$. To a solution of **20** (66 mg, 0.42 mmol) in $(CH_2Cl)_2$ (2 mL) at room temperature were added *i*-Pr₂NEt (0.72 mL, 4.16 mmol) and MOMCl (0.16 mL, 2.1 mmol). After stirring at room temperature for 3 h, the reaction mixture was quenched by addition of saturated aqueous NH4Cl solution. Products were extracted with EtOAc, and the organic layer was washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (1.6 g silica gel, 1:5 EtOAc/hexane as an eluent) to give 21a (81 mg, 96%) as a colorless syrup: $[\alpha]_D^{24}$ +47.0 (c 0.73, CHCl₃); IR $\nu_{\rm max}$ (neat) 2930, 2860, 1470, 1155, 1100 cm⁻¹; ¹H NMR δ 0.88 (3H, t, J=6.8 Hz), 1.21- 1.67 (10H, m), 2.53 (1H, dd, $J=2.7$, 4.9 Hz), 2.78 (1H, dd, $J=4.4$, 4.9 Hz), 2.98 (1H, ddd, J= 2.7 , 4.4, 7.1 Hz), 3.26 (1H, td, J=6.1, 7.1 Hz), 3.40 (3H, s), 4.66 and 4.88 (each 1H, 2d, $=$ 6.6 Hz); ¹³C NMR (75 MHz, CDCl₃) d 14.3, 22.8, 25.6, 29.5, 31.9, 32.5, 44.1, 54.9, 55.8, 78.2, 95.7; HRMS (FAB) m/z calcd for $C_{11}H_{22}O_3$ Na, $(M+Na)^+$ 225.1467, found: 225.1482.

4.3.4. (R)-2-{(R)-1-[(2-(Trimethylsilyl)ethoxy)methoxy]heptyl}oxirane (21b). To a solution of 20 (2.30 g, 14.3 mmol) in CH_2Cl_2 (65 mL) at room temperature were added i -Pr₂NEt (9.90 mL, 56.8 mmol) and SEMCl (5.00 mL, 28.4 mmol). After stirring at room temperature for 7 h, the reaction mixture was diluted with EtOAc and washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (150 g silica gel, 1:50–1:30 EtOAc/hexane as an eluent) to give 21b (4.20 g, 100%) as a colorless syrup: [α] $_D^{26}$ +52.0 (c 1.44, CHCl₃); IR $\nu_{\rm max}$ (neat) 2960, 2930, 2860, 1470, 1380, 1250, 1100 cm⁻¹; ¹H NMR δ 0.01 (9H, s), 0.84-0.98 (5H, m), 1.20-1.70 (10H, m), 2.52 (1H, dd, J=2.7, 4.9 Hz), 2.75 (1H, dd, $J=4.9, 4.9$ Hz), 2.96 (1H, ddd, J=2.7, 4.9, 6.8 Hz), 3.28 (1H, td, J=6.8, 7.1 Hz), 3.58 and 3.70 (each 1H, 2dt, $J=7.1$, 10.0 Hz), 4.71 and 4.88 (each 1H, 2d, J=6.8 Hz); ¹³C NMR δ -1.3, 14.2, 18.2, 22.7, 25.6, 29.5, 31.9, 32.5, 43.9, 54.9, 65.4, 77.8, 93.9; HRMS (FAB) m/z calcd for $C_{15}H_{33}O_3Si$, $(M+H)^+$ 289.2199, found: 289.2195.

4.3.5. tert-Butyl{(R)-1-[(R)-oxyran-2-yl]heptyloxy}diphenylsilane (21c). To a solution of 20 (257 mg, 1.63 mmol) in DMF (6.4 mL) at 0 °C were added imidazole (664 mg, 9.75 mmol) and TBDPSCI (1.25 mL, 4.88 mmol). After being stirred at room temperature for 6.5 h, the reaction mixture was diluted with EtOAc and washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (40 g silica gel, 1:100– 1:75 EtOAc/hexane as an eluent) to give 21c (644 mg, 100%) as a colorless syrup: $[\alpha]_D^{20}$ –17.5 (c 0.49, CHCl₃); IR $\nu_{\rm max}$ (neat) 2960, 2930, 2860, 1470, 1430, 1100, 1070, 930 cm $^{-1}$; 1 H NMR δ 0.82 (3H, t, $J=6.8$ Hz), 1.01–1.30 (17H, m), 1.45–1.52 (2H, m), 2.46 (1H, dd, J=2.8, 4.9 Hz), 2.71 (1H, dd, J=4.1, 4.9 Hz), 3.05 (1H, ddd, J=2.8, 4.1, 6.6 Hz), 3.35 (1H, td, J=6.1, 6.6 Hz), 7.34–7.45 (6H, m), 7.67–7.73 (4H, m); ^{13}C NMR δ 14.0, 19.4, 22.5, 24.8, 27.0, 29.0, 31.6, 34.7, 44.8, 55.7, 75.2, 127.4, 127.4, 129.5, 134.0, 134.3, 136.0; HRMS (FAB) m/z calcd for $C_{25}H_{37}O_2Si$, $(M+H)^+$ 397.2563, found: 397.2562.

4.3.6. $(R,E)-4-(Methoxymethoxy)dec-2-en-1-ol$ (22a). To a solution of 21a (581 mg, 2.87 mmol) in 1,4-dioxane (14 mL) at room temperature was added a solution of $HIO₄·2H₂O$ (1.50 g, 6.60 mmol) in H2O (2.9 mL). After being stirred at room temperature for 14 h, the reaction mixture was diluted with EtOAc and washed with brine, and dried. Removal of the solvent gave a crude aldehyde (571 mg). To a solution of the crude aldehyde (571 mg) in toluene (28 mL) was added $Ph_3P=CHCO_2Et$ (1.70 g, 4.90 mmol), and the resulting mixture was stirred at 60° C for 15 h. The reaction mixture was concentrated to give a residue, which was purified by column of silica gel (20 g, 1:100 EtOAc/hexane as an eluent) to afford a geometrical mixture ($E/Z = ca$. 6.5:1) of an ethyl ester (588 mg, 79%) as a yellow syrup. To a solution of the crude ethyl ester (588 mg, 2.28 mmol) in toluene (11 mL) was added at -78 °C under Ar dropwise DIBAL (0.99 M solution in toluene, 12.4 mL, 12.4 mmol), and the mixture was stirred at -78 °C for 20 min. The reaction mixture was quenched by addition of acetone (1.5 mL) at -78 °C. After stirring at

0 °C for 10 min, to the mixture was added $\text{Na}_2\text{SO}_4 \cdot 10 \text{H}_2\text{O}$ (2 g) and the whole mixture was further stirred at room temperature for 4 h. The insoluble material was removed by filtration through Celite, and the filtrate was concentrated to give a residue, which was purified by column chromatography (10 g silica gel, 1:8 EtOAc/hexane as an eluent) to afford first, Z-isomer of compound 22a (54 mg, 9% from **21a**) as a colorless syrup: [α] $_D^{23}$ +106.4 (*c* 0.58, CHCl $_3$); IR ν_{max} (neat) 3420, 2930, 2860, 1470, 1155, 1130 cm $^{-1}$; 1 H NMR δ 0.87 (3H, t, $J=6.6$ Hz), 1.19–1.69 (10H, m), 2.43 (1H, dd, J=4.4, 7.1 Hz), 3.37 (3H, s), 4.01 (1H, ddd, $J=1.2$, 6.1, 7.1 Hz), 4.26–4.42 (2H, m), 4.53 and 4.73 (each 1H, 2d, J=7.1 Hz), 5.36 (1H, ddd, J=1.2, 9.8, 11.0 Hz), 5.88 (1H, dddd, $J=1.0$, 6.1, 8.0, 11.0 Hz); ¹³C NMR δ 14.3, 22.8, 25.5, 29.4, 32.0, 35.6, 55.4, 58.3, 70.6, 93.6, 132.4, 132.7; HRMS (FAB) m/z calcd for $C_{12}H_{24}O_3$ Na, $(M+Na)^+$ 239.1624, found: 239.1618. Further elution afforded 22a (345 mg, 56% from 21a) as a colorless syrup: $[\alpha]_D^{23}$ +89.5 (c 0.51, CHCl₃); IR ν_{max} (neat) 3420, 2930, 2860, 1470, 1155, 1095 cm⁻¹; ¹H NMR δ 0.87 (3H, t, J=6.6 Hz), 1.22-1.68 (11H, m), 3.63 $(3H, s)$, 4.01 (1H, ddd, J=6.8, 6.8, 7.8 Hz), 4.14–4.17 (2H, m), 4.52 and 4.69 (each 1H, 2d, J=6.8 Hz), 5.55 (1H, bdd, J=7.8, 15.6 Hz), 5.81 (1H, btd, $J=5.4$, 15.6 Hz); ¹³C NMR δ 14.3, 22.8, 25.6, 29.4, 32.0, 35.8, 55.6, 63.1, 76.5, 93.9, 131.8, 132.1; HRMS (FAB) m/z calcd for C₁₂H₂₄O₃Na, $(M+Na)^+$ 239.1624, found: 239.1630. Anal. Calcd for C₁₂H₂₄O₃: C, 66.63; H, 11.18. Found: C, 66.60; H, 11.21.

4.3.7. (R,E)-4-{[(Trimethylsilyl)methoxy]ethoxy}dec-2-en-1-ol (22b). By the similar reaction conditions as described for the preparation of 22a from 21a, compound 21b (304 mg, 1.05 mmol) was converted into 22b (168 mg, 53%) and Z-isomer of 22b (22 mg, 7%). Data for Zisomer of **22b**: colorless syrup; [α] $_{{\rm D}}^{28}$ +104.0 (c 1.90, CHCl $_3$); IR $\nu_{\rm max}$ (neat) 3420, 2955, 2930, 2860, 1250, 1100, 1060 cm $^{-1}$; 1 H NMR δ 0.01 (9H, s), 0.84–1.00 (5H, m), 1.20–1.70 (10H, m), 2.47 (1H, br s), 3.53 and 3.71 (each 1H, 2dt, $J=7.1$, 10.0 Hz), 3.98 (1H, ddd, $J=1.0$, 6.1, 12.9 Hz), 4.32 (1H, ddd, J=1.0, 8.3, 12.9 Hz), 4.42 (1H, dt, J=6.3 and 10.0 Hz), 4.61 and 4.74 (each 1H, 2d, J=7.1 Hz), 5.35 (1H, dddd, J=1.0, 1.0, 10.0, 11.0 Hz), 5.89 (1H, ddd, J=6.1, 8.3, 11.0 Hz); ¹³C NMR δ -1.4, 14.2,18.1, 22.7, 25.5, 29.4, 31.9, 35.6, 58.2, 65.1, 70.7, 91.7,132.4,132.7; HRMS (FAB) m/z calcd for C₁₆H₃₅O₃Si, $(M+H)^+$ 303.2356, found: 303.2353. Data for **22b**: colorless syrup; $[\alpha]_D^{27}$ +86.4 (*c* 2.43, CHCl₃); IR ν_{max} (neat) 3400, 2930, 2860, 1460, 1380, 1250, 1100, 1060 cm $^{-1};$ 1 H NMR δ 0.02 (9H, s), 0.85–0.96 (5H, m), 1.20–1.70 (11H, m), 3.51 and 3.73 (each 1H, 2dt, J=7.1, 10.0 Hz), 4.04 (1H, dt, J=5.9, 7.8 Hz), 4.15 (2H, dd, J=1.2, 5.3 Hz), 4.60 and 4.69 (each 1H, 2d, J=7.1 Hz), 5.55 (1H, dddd, J=1.2, 1.2, 7.8, 15.6 Hz), 5.81 (1H, td, J=5.3, 15.6 Hz); ¹³C $NMR\delta - 1.3, 14.2, 18.3, 22.8, 25.6, 29.4, 32.0, 35.8, 63.1, 65.2, 76.2, 92.1,$ 131.9, 132.0; HRMS (FAB) m/z calcd for C₁₆H₃₅O₃Si, $(M+H)^+$ 303.2356, found: 303.2356.

4.3.8. (R,E)-4-(tert-Butyldiphenylsilyloxy)dec-2-en-1-ol (22c). By the similar reaction conditions as described for the preparation of 22a from 21a, compound $21c(1.00 g, 2.52 mmol)$ was converted into 22c (692 mg, 67%) and its Z-isomer (49 mg, 5%). Data for Z-isomer of **22c**: colorless syrup; [α] $_{{\rm D}}^{19}$ -24.4 (*c* 0.595, CHCl₃); IR $\nu_{\rm max}$ (neat) 3300, 2960, 2930, 2860, 1470, 1430, 1100, 1080 cm $^{-1}$; 1 H NMR δ 0.86 $(3H, t, J=6.7 Hz)$, 1.05 (9H, s), 1.18–1.29 (9H, m), 1.39–1.66 (2H, m) 3.58–3.69 (2H, m), 4.35 (1H, td, J=7.3, 8.7 Hz), 5.33 (1H, td, J₂₋₁=6.8, 11.2 Hz), 5.49 (1H, dd, J=8.7, 11.2 Hz), 7.34–7.47(6H, m), 7.66–7.71 (4H, m); ¹³C NMR (75 MHz, CDCl₃) δ 14.1, 19.3, 22.6, 24.8, 26.9, 29.2, 31.8, 38.2, 58.6, 69.4, 127.4, 127.5, 128.1, 129.6, 129.7, 134.2, 134.3, 135.9,136.1; HRMS (FAB) m/z calcd for C₂₆H₃₉O₂Si, $(M+H)^+$ 411.2719, found: 411.2713. Data for **22c**: colorless syrup; $[\alpha]_D^{21}$ +5.9 (*c* 0.66, MeOH); IR v_{max} (neat) 3300, 2960, 2930, 2860, 1470, 1430, 1100, 1080, 970 cm⁻¹; ¹H NMR δ 0.86 (3H, t, J=6.9 Hz), 1.07 (9H, s), 1.11- 1.59 (11H, m), 3.90 (2H, dd, J=1.2, 5.6 Hz), 4.18 (1H, dt, J=6.1, 6.3 Hz), 5.40 (1H, td, J=5.6, 15.4 Hz), 5.56 (1H, tdd, J=1.2, 6.1, 15.4 Hz), 7.33– 7.54 (6H, m), 7.65–7.70 (4H, m); ¹³C NMR (75 MHz, CDCl₃) δ 14.1, 19.3, 22.6, 24.7, 27.0, 29.2, 31.8, 37.9, 63.1, 73.9, 127.3, 127.5, 129.1, 129.4, 129.5, 134.4, 134.7, 135.9, 136.1; HRMS (FAB) m/z calcd for $C_{26}H_{38}O_2SiNa$, $(M+Na)^+$ 433.2539, found: 433.2550.

4.3.9. (R,E)-7-(Methoxymethoxy)trideca-1,5-diene (23a). To a solution of 22a (339 mg, 1.57 mmol) in $\mathrm{CH}_2\mathrm{Cl}_2$ (8 mL) at 0 °C were added Et3N (0.440 mL, 3.16 mmol) and MsCl (0.220 mL, 2.84 mmol). After stirring at 0° C for 30 min, the reaction was quenched by addition of 1 M aqueous HCl solution (2 mL) at 0 \degree C. The resulting mixture was diluted with EtOAc and washed successively with 1 M aqueous HCl solution, saturated aqueous $NAHCO₃$ solution and brine, and dried. Removal of the solvent gave a crude mesylate (502 mg). To a solution of the crude mesylate (502 mg) in THF (16 mL) under Ar at room temperature was added allylmagnesium chloride (2.0 M solution in THF, 2.75 mL, 5.50 mmol), and the mixture was stirred at 50 $\,^{\circ}$ C for 10 h. The reaction was quenched by addition of 1 M aqueous HCl solution (5 mL) at 0° C. The mixture was diluted with EtOAc and washed successively with 1 M aqueous HCl solution, saturated aqueous NaHCO $_3$ solution and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (10 g silica gel, 1:15 EtOAc/hexane as an eluent) to afford 23a (307 mg, 81%) as a colorless syrup: $[\alpha]_D^{23}$ +99.1 (c 0.92, CHCl₃); IR ν_{max} (neat) 2930, 2860, 1470, 1115, 1095, 1040 cm $^{-1}$; 1 H NMR δ 0.88 $(3H, t, J=7.1 Hz)$, 1.21–1.66 (10H, m), 2.12–2.15 (4H, m), 3.35 (3H, s), 3.92 (1H, td, J=6.3, 8.3 Hz), 4.48 and 4.71 (each 1H, 2d, J=6.6 Hz), 4.96 (1H, d, J=10.2 Hz), 5.01 (1H, d, J=18.5 Hz), 5.26 (1H, dd, J=8.3, 15.3 Hz), 5.60 (1H, td, J=6.3, 15.3 Hz), 5.72–5.86 (1H, m); ¹³C NMR d 14.3, 22.8, 25.7, 29.4, 31.8, 32.0, 33.6, 35.9, 55.5, 76.9, 93.4, 115.0, 130.8, 133.7, 138.3; HRMS (FAB) m/z calcd for C₁₅H₂₈O₂Na, (M+Na)⁺ 263.1987, found: 263.1982.

4.3.10. (R,E)-Trimethyl{[2-(trideca-1,5-dien-7-yloxy)ethoxy]methyl} silane (23b). By the similar reaction conditions as described for the preparation of 23a from 22a, compound 22b (954 mg, 3.16 mmol) was converted into 23b (691 mg, 67%): colorless syrup; $\lbrack \alpha \rbrack^{27}_D + 98.4$ (c 0.74, CHCl₃); IR ν_{max} (neat) 2960, 2930, 2860, 1640, 1380, 1250, 1100, 1060 cm⁻¹; ¹H NMR δ 0.02 (9H, s), 0.85-0.96 (5H, m), 1.20-1.65 (10H, m), 2.12–2.15 (4H, m), 3.49 and 3.73 (each 1H, 2dt, $I=6.6$, 9.5 Hz), 3.94 (1H, dt, $J=6.3$, 6.8 Hz), 4.57 and 4.70 (each 1H, 2d, $J=6.8$ Hz), 4.96 (1H, dd, J=2.0, 11.0 Hz), 5.01 (1H, dd, J=2.0, 18.1 Hz), 5.26 (1H, dd, J=8.3, 15.4 Hz), 5.55–5.66 (1H, m), 5.72–5.86 (1H, m); 13 C NMR δ -1.3, 14.2, 18.3, 22.8, 25.7, 29.4, 31.8, 32.0, 33.6, 35.9, 65.1, 76.8, 91.7, 115.0, 130.9, 133.4, 138.2; HRMS (FAB) m/z calcd for $C_{19}H_{39}O_2Si$, $(M+H)^+$ 327.2719, found: 327.2712.

4.3.11. (R,E)-tert-Butyldiphenyl(trideca-1,5-dien-7-yloxy)silane (23c). By the similar reaction conditions as described for the preparation of 23a from 22a, compound 22c (217 mg, 0.527 mmol) was converted into 23c (192 mg, 84%): colorless syrup; $\lbrack \alpha \rbrack^{26}_{{\rm D}}+21.8$ (c 0.75, CHCl₃); IR ν_{max} (neat) 2960, 2930, 2860, 1470, 1430, 1100, 1070, 970 cm⁻¹; ¹H NMR δ 0.85 (3H, t, J=6.8 Hz), 1.05 (9H, s), 1.10-1.55 (10H, m), 1.97-1.99 (4H, m), 4.06 (1H, ddt, J_{7-6} =1.0, 7.1, 7.1 Hz), 4.91 (1H, ddd, J=1.0, 2.2, 10.1 Hz), 4.96 (1H, ddd, J=1.0, 2.2, 17.1 Hz), 5.23 (1H, td, J=5.6, 15.4 Hz), 5.39 (1H, dd, J=7.1, 15.4 Hz), 5.67-5.80 (1H, m), 7.31–7.44 (6H, m), 7.64–7.70 (4H, m); ¹³C NMR δ 14.1, 19.3, 22.6, 24.8, 27.1, 29.2, 31.5, 31.8, 33.3, 38.1, 74.7, 114.5, 127.2, 127.4, 129.3, 129.4, 130.1, 133.4, 134.71, 134.74, 136.0, 136.1, 138.3; HRMS (FAB) m/z calcd for C₂₉H₄₃OSi, $(M+H)^+$ 435.3083, found: 435.3098.

4.3.12. (R, E) -7-(Methoxymethoxy)tridec-5-en-1-ol $(24a)$. To neat 23a (135 mg, 0.56 mmol) under Ar at room temperature was added a solution of 9-BBN (0.5 M in THF, 9.0 mL, 4.5 mmol), and the resulting mixture was sonicated (47 kHz, 60 W) in a water bath at ambient temperature for 2 h. To the reaction mixture at 0° C were added H₂O (9 mL) and NaBO₃ $4H₂O$ (2.58 g, 16.8 mmol), and the mixture was further stirred at $0 °C$ for 17 h. The reaction mixture was diluted with EtOAc and washed with brine, and dried. Removal

of the solvent left a residue, which was purified by column chromatography (15 g silica gel, 1:5 EtOAc/hexane as an eluent) to afford **24a** (124 mg, 86%) as a colorless syrup: [α] $_D^{27}$ +95.2 (c 1.05, CHCl $_3$); IR $\rm \nu_{max}$ (neat) 3400, 2930, 2860, 1470, 1115, 1095, 1040 cm $^{-1};\,{}^{1}$ H NMR δ 0.87 (3H, t, J=6.3 Hz), 1.22-1.70 (15H, m), 2.07 (2H, dt, J=6.8, 7.1 Hz), 3.35 (3H, s), 3.64 (2H, t, J=6.6 Hz), 3.92 (1H, td, J=6.6, 8.3 Hz), 4.49 and 4.71 (each 1H, 2d, J=6.8 Hz), 5.26 (1H, dd, J=8.3, 15.4 Hz), 5.60 (1H, td, J=6.8, 15.4 Hz); ¹³C NMR δ 14.3, 22.8, 25.5, 25.7, 29.4, 32.0, 32.1, 32.4, 35.9, 55.5, 62.9, 77.0, 93.4, 130.7, 134.1; HRMS (FAB) m/z calcd for C₁₅H₃₀O₃Na, $(M+Na)^{+}$ 281.2093, found: 281.2094.

4.3.13. (R,E)-7-{[(Trimethylsilyl)methoxy]ethoxy}tridec-5-en-1-ol (24b). By the similar reaction conditions as described for the preparation of 24a from 23a, compound 23b (993 mg, 3.04 mmol) was converted into **24b** (1.17 g, 100%): colorless syrup; $[\alpha]_D^{20}$ +96.1 (c 1.45, CHCl₃); IR ν_{max} (neat) 3420, 2930, 2860, 1455, 1380, 1250, 1100, 1055 cm⁻¹; ¹H NMR δ 0.02 (9H, s), 0.85-0.99 (5H, m), 1.23-1.62 (15H, m), 2.07 (2H, dtd, J=1.5, 6.1, 7.1 Hz), 3.50 (1H, dt, J=7.3, 9.5 Hz), 3.64 (2H, t, J=6.3 Hz), 3.73 (1H, td, J=7.3, 9.5 Hz), 3.94 (1H, td, $J=5.9$, 8.3 Hz), 4.58 and 4.70 (each 1H, 2d, $J=6.8$ Hz), 5.26 (1H, tdd, J = 1.5, 8.3, 15.4 Hz), 5.60 (1H, td, J = 7.1, 15.4 Hz); ¹³C NMR δ – 1.3, 14.2, 18.3, 22.8, 25.5, 25.7, 29.4, 32.0, 32.1, 32.4, 35.9, 62.9, 65.1, 76.8, 91.7, 130.8, 133.9; HRMS (FAB) m/z calcd for C₁₉H₄₁O₃Si, $(M+H)^+$ 345.2825, found: 345.2808.

4.3.14. (R,E)-7-{[(Trimethylsilyl)methoxy]ethoxy}tridec-5-en-1-ol ($24c$). By the similar reaction conditions as described for the preparation of $24a$ from $23a$, compound $23c$ (56 mg, 0.014 mmol) was converted into **24c** (53 mg, 90%): colorless syrup; [α] $^{26}_{\rm D}$ +14.3 (*c* 0.26, CHCl₃); IR v_{max} (neat) 3350, 2960, 2930, 2860, 1470, 1430, 1100 cm⁻¹; ¹H NMR δ 0.83 (3H, t, J=7.0 Hz), 1.03 (9H, s), 1.15-1.58 $(15H, m)$, 1.88 (2H, tdd, J=1.0, 6.8, 6.8 Hz), 3.56 (2H, t, J=6.4 Hz), 4.05 $(dt, J=6.8, 6.8 Hz)$, 5.18 (1H, tdd, J=1.0, 6.8, 15.5 Hz), 5.36 (1H, tdd, J=1.0, 6.8, 15.5 Hz), 7.30–7.41 (6H, m), 7.63–7.67 (4H, m); ¹³C NMR d 14.0,19.3, 22.6, 24.8, 25.2, 27.1, 29.2, 31.8, 32.2, 38.1, 62.8, 74.7,127.2, 127.4, 129.2, 129.4, 130.6, 133.3, 134.7, 134.8, 135.9, 136.0; HRMS (FAB) m/z calcd for C₂₉H₄₅O₂Si, $(M+H)^+$ 452.3189, found: 452.3198.

4.3.15. (R,E)-1-Iodo-7-(methoxymethoxy)tridec-5-ene ($5a$). To a solution of 24a (172 mg, 0.67 mmol) in CH₃CN (5.0 mL) and Et₂O (1.7 mL) at 0 °C were added imidazole $(273 \text{ mg}, 4.01 \text{ mmol})$, Ph₃P (526 mg, 2.01 mmol), and I_2 (382 mg, 3.01 mmol). After stirring at room temperature for 13 h, the reaction mixture was diluted with hexane (5 mL) and the insoluble material was removed by filtration through a pad of silica gel. The filtrate was concentrated to give a residue, which was purified by column chromatography (20 g silica gel, toluene as an eluent) to afford 5a (215 mg, 87%) as a colorless syrup: [α] $_{{\rm D}}^{\rm 26}$ +69.9 (c 3.71, CHCl₃); IR $\nu_{\rm max}$ (neat) 2915, 2855, 1455, 1150, 1095 cm⁻¹; ¹H NMR δ 0.87 (3H, t, J=6.8 Hz), 1.24-1.54 (12H, m), 1.82 (2H, m), 2.07 (2H, dt, $J=6.6$, 6.8 Hz), 3.18 (2H, t, J=7.1 Hz), 3.36 (3H, s), 3.92 (2H, td, J=6.6, 8.0 Hz), 4.50 and 4.70 (each 1H, 2d, J=6.6 Hz), 5.27 (1H, dd, J=8.0, 15.3 Hz), 5.58 (1H, ddd, J=6.6, 6.6, 15.3 Hz); ¹³C NMR δ 6.8, 14.3, 22.8, 25.7, 29.4, 30.1, 31.3, 32.0, 33.1, 35.9, 55.5, 76.9, 93.5, 131.1, 133.5; HRMS (FAB) m/z calcd for $C_{15}H_{29}IO_2$ Na, $(M+Na)^+$ 391.1111, found: 391.1108. Anal. Calcd for C15H29IO2: C, 48.92; H, 7.94. Found: C, 48.80; H, 7.94.

4.3.16. (R,E)-1-Iodo-7-{[(trimethylsilyl)methoxy]ethoxy}tridec-5-ene $(5b)$. By the similar reaction conditions as described for the preparation of 5a from 24a, compound 24b (28 mg, 0.080 mmol) was converted into **5b** (31 mg, 84%): colorless syrup; [α] $_{{\rm D}}^{28}$ +70.8 (*c* 1.43, CHCl₃); IR ν_{max} (neat) 2955, 2930, 2855, 1460, 1380, 1250, 1100 cm⁻¹; ¹H NMR δ 0.02 (9H, s), 0.85-0.96 (5H, m), 1.24-1.61 $(12H, m)$, 1.82 $(2H, tt, J=7.1, 7.1 Hz)$, 2.06 $(2H, dt, J=6.8, 7.1 Hz)$, 3.17 (2H, t, J=7.1 Hz), 3.49 and 3.75 (each 1H, 2td, J=7.1, 9.8 Hz), 3.95 (1H, td, J=6.6, 8.1 Hz), 4.58 and 4.69 (each 1H, 2d, J=6.8 Hz), 5.26 (1H, tdd, $J=1.0$, 8.1, 15.3 Hz), 5.58 (1H, ddd, $J=6.8$, 6.8, 15.4 Hz); ^{13}C NMR $δ$ - 1.3, 6.8, 14.3, 18.3, 22.8, 25.7, 25.7, 29.4, 30.1, 31.2, 32.0, 33.1, 35.9, 65.1, 76.7, 91.7, 131.2, 133.3; HRMS (FAB) m/z calcd for $C_{19}H_{40}O_{2}$ ISi, $(M+H)^{+}$ 455.1843, found: 455.1853.

4.3.17. Determination of optical purities of 5a, 5b, and 24c. Starting from dimethyl *L*-tartrate, enantiomers of $5a$ (ent- $5a$), $5b$ (ent- $5b$), and 24c (ent-24c) were synthesized by the similar reaction procedures. Optical purities of 5a, ent-5a, 5b, ent-5b, 24c, and ent-24c were determined to be all >99% ee by chiral HPLC analyses. Analytical conditions for 5a and 24c: [DAICEL CHIRALCEL OD-H, 4.6 mm ID \times 250 mm, *i*-PrOH/hexane=1:500, flow rate=1.0 mL/ min, UV (254 nm) detection]; retention time for 5a, 5.8 min; for ent-5a, 5.1 min; retention rime for 24c, 34.6 min; for ent-24c, 31.8 min. Analytical conditions for 5b: [DAICEL CHIRALCEL OD-H, 4.6 mm ID \times 250 mm, *i*-PrOH/hexane=1:1000, flow rate=1.0 mL/ min, UV (254 nm) detection]; retention time for 5b, 7.0 min; for ent-5b, 5.0 min.

4.4. Coupling reactions of the left-half segment with the right-half segment and total synthesis of mycestericin A

4.4.1. Methyl (4R,5S)-4-[(1R,3E,9E,11R)-1,11-bis(methoxymethoxy)- 3,9-heptadecadiene-1-yl]-2,2-dimethyl-5-[(2,2,2-trichloroacetyl) amino]-1,3-dioxane-5-carboxylate (25). To a solution of 5a (30 mg, 0.082 mmol) in THF (0.2 mL) was added tert-BuLi (1.53 M solution in pentane, 0.28 mL, 0.43 mmol) at -78 °C under Ar, and the mixture was stirred for 15 min at -78 °C. To this mixture at -78 °C was added a THF solution of $ZnCl₂$ (1.0 M, 0.13 mL, 0.13 mmol), and the cooling bath was removed. After stirring at ambient temperature for 30 min, the resulting colorless solution of Zn reagent was added to a solution of $2(11 \text{ mg}, 0.019 \text{ mmol})$ and $Pd(PPh_3)_4(2.3 \text{ mg}, 0.002 \text{ mmol})$ in THF (0.25 mL) and benzene (0.25 mL) via a cannula at room temperature. After stirring at room temperature for 2 h, the reaction was quenched by addition of saturated aqueous NH4Cl solution (2 mL) at 0 \degree C, and products were extracted with Et₂O. The organic layer was washed successively with saturated aqueous NH4Cl solution, saturated aqueous NaHCO₃ solution and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (1.3 g silica gel, 1:20 EtOAc/toluene as an eluent) to afford 25 (11.3 mg, 86%) as a colorless syrup: [α] $_{{\rm D}}^{25}$ +82.6 (c 0.21, CHCl3); IR $\nu_{\rm max}$ (neat) 3375, 2930, 2855, 1740, 1700, 1520, 1380, 1260, 1200 cm $^{-1}$; $^1\mathrm{H}$ NMR δ 0.88 (3H, t, J=6.8 Hz), 1.24–1.58 (14H, m), 1.45 and 1.50 (each 3H, 2s),1.95–2.09 (4H, m), 2.39–2.46 (2H, m), 3.36 and 3.45 (each 3H, 2s), 3.70 (1H, ddd, J=1.0, 6.1, 8.3 Hz), 3.77 (3H, s), 3.92 (1H, dt, J=6.6, 7.8 Hz), 4.08 (1H, d, J=1 Hz), 4.25 (1H, d, J=12.2 Hz), 4.49 (1H, d, J=6.6 Hz), 4.52 (1H, d, J=12.2 Hz), 4.71 (1H, d, J=6.6 Hz), 4.81 and 4.85 (each 1H, 2d, J=7.1 Hz), 5.18–5.30 (2H, m), 5.50 (1H, ddd, J=6.8, 6.8, 15.4 Hz), 5.60 (1H, ddd, J=6.8, 6.8, 15.4 Hz), 8.54 (1H, br s); ¹³C NMR d 14.3, 18.9, 22.8, 25.7, 28.9, 29.1, 29.1, 29.4, 32.1, 32.3, 32.7, 34.1, 35.9, 53.0, 55.5, 56.8, 62.0, 62.7, 66.7, 71.0, 76.9, 77.0, 93.4, 96.5, 100.1, 124.5, 130.5, 134.3, 135.2, 163.7, 169.2; HRMS (FAB) m/z calcd for $C_{31}H_{52}Cl_{3}NO_{9}Na$, $(M+Na)^+$ 710.2605, found: 710.2600.

4.4.2. N-{(3S,4R,5R)-4-(Acetyloxy)-3-[(acetoxy)methyl]tetrahydro-2 oxo-5-[(2E,8E,10R)-10-[[2-(trimethylsilyl)ethoxy]methoxy]-2,8-hexadecadien-1-yll-3-furanyl}-acetamide (31). To a solution of 5b (226 mg, 0.050 mmol) in THF (4.0 mL) was added tert-BuLi (1.53 M solution in pentane, 0.65 mL, 1.00 mmol) at -78 $^{\circ}$ C under Ar, and the mixture was stirred for 15 min at -78 °C. To this mixture at -78 °C was added $ZnCl₂$ (1.0 M solution in THF, 0.50 mL, 0.50 mmol), and the cooling bath was removed. After stirring at ambient temperature for 30 min, the resulting colorless solution of Zn reagent was added to a solution of **3** (56 mg, 0.13 mmol) and $Pd(PPh_3)_4$ (43.7 mg, 0.038 mmol) in THF (5.0 mL) via a cannula at room temperature. After stirring at room temperature for 1.5 h, the reaction was quenched by addition of saturated aqueous NH4Cl solution (15 mL) at 0 $^{\circ}$ C, and products were extracted with Et $_{2}$ O. The organic layer was washed successively with saturated aqueous NH4Cl solution, saturated aqueous NaHCO₃ solution and brine, and dried. Removal of the solvent left a residue, which was roughly purified by column chromatography (3 g silica gel, 1:1 EtOAc/hexane as an eluent) to afford crude coupling product. The crude product was dissolved in pyridine (2.0 mL) and Ac₂O (1.0 mL) and the resulting solution was stirred at room temperature for 5 h. The reaction mixture was concentrated to give a residue, which was purified by column chromatography (3 g silica gel, 1:2–1:1 EtOAc/hexane as an eluent) to afford 31 (35.1 mg, 44%) as a colorless syrup: [α] $_{{\rm D}}^{\rm 24}$ +95.9 (c 1.14, CHCl3); IR $\nu_{\rm max}$ (neat) 3300, 2950, 2930, 2860, 1790, 1755, 1750, 1680, 1370, 1230, 1055 cm⁻¹; ¹H NMR δ 0.02 (9H, s), 0.83-0.96 (5H, m), 1.20-1.60 (14H, m), 1.92–2.15 (4H, m), 2.02, 2.04 and 2.10 (each 3 H, 3s), 2.31–2.46 $(2H, m)$, 3.50 and 3.72 (each 1H, 2dt, J=6.6, 9.8 Hz), 3.94 (1H, dt, J=7.1, 7.1 Hz), 4.51 (2H, s), 4.58 and 4.70 (each 1H, 2d, $J=6.8$ Hz), 4.68-4.74 $(1H, m)$, 5.23 (1H, dd, J=6.1, 15.3 Hz), 5.40 (1H, ddd, J=6.6, 6.6, 15.6 Hz), 5.51–5.63 (2H, m), 5.79 (1H, d, J=4.2 Hz), 6.00 (1H, br s); ¹³C NMR $δ$ -1.3, 14.2, 18.2, 20.5, 20.7, 22.8, 22.9, 25.7, 28.8, 29.3, 32.0, 32.2, 32.3, 32.5, 35.9, 62.8, 72.0, 76.7, 81.7, 91.5, 123.3, 130.4, 134.2, 135.1, 169.0, 169.6, 170.3, 172.6; HRMS (FAB) m/z calcd for $C_{33}H_{58}NO_9Si$, $(M+H)^+$ 640.3881, found: 640.3878.

4.4.3. N-{(3S,4R,5R)-4-(Acetyloxy)-5-[(2E,8E,10R)-10-(acetyloxy)- 2,8-hexadecadien-1-yl]-3-[(acetyloxy)methyl]tetrahydro-2-oxo-3 furanyl}-acetamide (tetraacetyl mycestericin A γ -lactone) (28b). To a mixture of 31 (2.8 mg, 4.4μ mol) and MS4A (50 mg) at room temperature under Ar was added Bu4NF (57.3 mg, 0.22 mmol) in N,N-dimethyl propylene urea (DMPU, 1.0 mL), and the resulting mixture was stirred at 80 \degree C for 12 h. After cooling, to the reaction mixture at room temperature were added pyridine (1.0 mL) and $Ac₂O$ (0.5 mL). After stirring at room temperature for 15 h, the reaction mixture was poured into $H₂O$ (2 mL), and products were extracted with EtOAc. The organic layer was washed with brine, and dried. Removal of the solvent left a residue, which was purified by preparative TLC (4:1 EtOAc/hexane as an eluent) to afford 28b (0.9 mg, 38%) as a colorless syrup: $[\alpha]_D^{21}$ +58.2 (c 0.41, CHCl₃); IR v_{max} (neat) 3350, 2920, 2855, 1790, 1750, 1725, 1680, 1540, 1460, 1375, 1240, 1030 cm⁻¹; ¹H NMR δ 0.87 (3H, t, J=7.0 Hz), 1.22-1.40 (12H, m), 1.50–1.66 (2H, m), 1.99–2.05 (4H, m), 2.02, 2.03, 2.05, and 2.10 (each 3 H, 4s), 2.29–2.49 (2H, m), 4.50 and 4.52 (each 1H, 2d, $J=11.5$ Hz), 4.71 (1H, ddd, J=4.4, 8.0, 8.0 Hz), 5.17 (1H, dt, J=6.8, 6.8 Hz), 5.32–5.44 (2H, m), 5.51–5.71 (2H, m), 5.79 (1H, d, J=4.4 Hz), 6.01 (1H, br s); 13C NMR d 14.0, 20.3, 20.5, 21.4, 22.5, 22.7, 25.2, 28.5, 28.7, 29.0, 31.7, 32.0, 32.2, 32.3, 34.5, 62.6, 62.7, 71.8, 75.0, 81.6, 123.2, 128.6, 134.0, 135.0, 168.8, 169.3, 170.2, 170.4, 172.4; HRMS (FAB) m/z calcd for C₂₉H₄₆NO₉, (M+H)⁺ 552.3172, found: 552.3181. The spectral data were fully identical with those reported for the authentic sample derived from natural mycestericin A.^{1a}

4.4.4. N-{(3S,4R,5R)-4-(Acetyloxy)-3-[(acetoxy)methyl]tetrahydro-2 oxo-5-[(2E,8E,10R)-10-(tert-butyldimethylsilyloxy)-2,8-hexadecadien-1-yl]-3-furanyl}-acetamide (32). To neat 23c (25 mg, 0.057 mmol) under Ar at room temperature was added a solution of 9-BBN (0.5 M in THF, 0.91 mL, 0.46 mmol), and the resulting mixture was sonicated (47 kHz, 60 W) in a water bath at ambient temperature for 1 h. To the reaction mixture at 0 $^{\circ}$ C was added H₂O (0.12 mL), and the mixture was stirred at room temperature for 10 min. The resulting organoborane solution was added to a solution of 3 (6.1 mg, 0.014 mmol) and AsPh₃ (0.9 mg, 2.8 μ mol) in DMF (1.0 mL) via a cannula at room temperature. To this mixture at room temperature were added K_2CO_3 (7.9 mg, 0.057 mmol) and PdCl₂(dppf) \cdot CH₂Cl₂ (2.3 mg, 2.8 µmol), and the whole mixture was stirred at room temperature for 1.5 h. The reaction mixture was diluted with saturated aqueous NH_4Cl solution at $0 °C$, and

products were extracted with EtOAc. The organic layer was washed successively with saturated aqueous NH4Cl solution, water and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (0.3 g silica gel, 1:3 EtOAc/ hexane as an eluent) to afford 32 (9.0 mg, 87%) as a colorless syrup: $[\alpha]_D^{25}$ +47.2 (c 0.94, CHCl₃); IR ν_{max} (neat) 3350, 2950, 2930, 2860, 1790, 1755, 1750, 1680, 1540, 1460, 1430, 1370, 1230, 1190, 1110, $1055, 1030$ cm⁻¹; ¹H NMR δ 0.84 (3H, t, J=6.6 Hz), 1.04 (9H, s), 1.15-1.62 (14H, m), 1.83–2.11 (4H, m), 2.03, 2.04, and 2.09 (each 3H, 3s), 2.34–2.43 (2H, m), 4.05 (1H, ddt, $J=1.0$, 6.8, 6.8 Hz), 4.49 and 4.53 (each 1H, 2d, J=13.4 Hz), 4.71 (1H, ddd, J=4.6, 8.3, 8.3 Hz), 5.20 (1H, dddd, J=1.0, 6.6, 6.6, 15.4 Hz), 5.32–5.43 (2H, m), 5.54 (1H, dddd, $J=1.0$, 6.6, 6.6, 15.4 Hz), 5.79 (1H, d, $J=4.6$ Hz), 6.03 (1H, br s), 7.30– 7.42 (6H, m), 7.63-7.68 (4H, m); 13 C NMR δ 14.0, 19.3, 20.3, 20.5, 22.6, 22.8, 24.8, 27.1, 28.6, 28.7, 29.3, 31.8, 31.9, 32.2, 32.4, 38.1, 62.6, 62.7, 71.9, 74.7, 81.6, 123.1, 127.2, 127.3, 129.2, 129.4, 130.8, 133.0, 134.7, 135.1, 135.9, 136.0, 168.0, 169.3, 170.1, 172.4; HRMS (FAB) m/z calcd for $C_{43}H_{62}NO_8Si$, $(M+H)^+$ 748.4244, found: 748.4251.

4.4.5. Tetraacetyl mycestericin A γ -lactone (28b) from 32. To a solution of 32 (3.7 mg, 4.9 μ mol) in THF (0.7 mL) at room temperature was added Bu₄NF (1.0 M solution in THF, 90 μ L, 90 μ mol), and the mixture was stirred at $65 °C$ for 7 h. After cooling, to the resulting mixture were added Ac_2O (0.5 mL) and pyridine (1.0 mL). After stirring at room temperature for 15 h, the reaction mixture was diluted with H_2O at $0 °C$, and products were extracted with EtOAc. The organic layer was washed successively with H_2O and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (0.3 g silica gel, 1:2–1:1.5 EtOAc/hexane as an eluent) to give **26b** (1.7 mg, 63%) as a colorless syrup. The $[\alpha]_D$ value and spectral (IR, ${}^{1}H$ and ${}^{13}C$ NMR, and MS) data were fully identical with those of 28b obtained from 31.

4.4.6. Mycestericin A (1) . To a solution of **28b** $(6.8 \text{ mg}, 1.2 \text{ µmol})$ in MeOH (0.5 mL) at room temperature was added 15% aqueous NaOH solution (0.5 mL). The mixture was stirred at 70 \degree C for 3 h, and then neutralized with IRC-76 resin (H^+ form) at room temperature. Insoluble materials were removed by filtration and the filtrate was concentrated to give a residue, which was purified by column chromatography [0.3 g silica gel, 1:5-2:5 MeOH/CHCl₃ (saturated with H₂O) as an eluent] to afford mycestericin $A(1)(4.6$ mg, 93%) as an amorphous solid: $[\alpha]_D^{21}$ –8.6 (c 0.45, MeOH); IR ν_{max} (KBr disk) 3400, 3270, 2920, 2855, 1630, 1460, 1400, 1050, 965 cm⁻¹; ¹H NMR (CD₃OD) δ 0.89 (3H, t, J=7.1 Hz), 1.25–1.60 (14H, m), 1.93–2.10 (4H, m), 2.26 (2H, t, J=6.8 Hz), 3.77-3.95 (4H, m), 4.00 (1H, d, J=11.0 Hz), 5.34–5.47 (2H, m), 5.53 (1H, ddd, J=6.8, 6.8, 15.3 Hz), 5.59 (1H, ddd, J=6.8, 6.8 15.3 Hz); ¹³C NMR (CD₃OD) δ 14.4, 23.6, 26.6, 29.97, 30.0, 30.4, 33.0, 33.1, 33.6, 38.5, 38.6, 65.2, 70.5, 71.2, 73.66, 73.74, 126.9, 132.3, 134.57, 134.64, 173.5; HRMS (FAB) m/z calcd for C₂₁H₄₀NO₆, $(M+H)^+$ 402.2856, found: 402.2857. The spectral data were fully identical with those reported for natural mycestericin A^{1a}

4.5. Synthesis of 14-epi-mycestericin A

4.5.1. (S,E)-tert-Butyldiphenyl(trideca-1,5-dien-7-yloxy)silane (ent-23c). The same reaction sequence for the preparation of 23c from diisopropyl D-tartrate was applied to dimethyl L-tartrate to provide *ent*-23c: $[\alpha]_D^{26}$ –21.2 (c 0.55, CHCl₃). The spectral data were fully identical with those of 23c.

4.5.2. N-{(3S,4R,5R)-4-(Acetyloxy)-3-[(acetoxy)methyl]tetrahydro-2 oxo-5-[(2E,8E,10S)-10-(tert-butyldimethylsilyloxy)-2,8-hexadecadien-1-yl]-3-furanyl}-acetamide (33). Coupling reaction of ent-23c (56 mg, 0.13 mmol) with 3 (15 mg, 0.034 mmol) by the similar reaction conditions as described for the reaction of 23c with 3 afforded 33 (19 mg, 77%) as a colorless syrup: $[\alpha]_D^{25}$ +26.6 (c 0.95,

CHCl₃); IR v_{max} (neat) 3350, 2950, 2930, 2860, 1790, 1755, 1750, 1680, 1540, 1460, 1430, 1370, 1230, 1190, 1110, 1055, 1030 cm⁻¹; ¹H NMR δ 0.84 (3H, t, J=6.6 Hz), 1.04 (9H, s), 1.15-1.62 (14H, m), 1.83-2.11 (4H, m), 2.03, 2.04 and 2.09 (each 3H, 3s), 2.29–2.48 (2H, m), 4.05 (ddt, 1H, H-14, $J=1$, 6.8, 6.8 Hz), 4.49 and 4.53 (each 1H, 2d, J=13.4 Hz), 4.70 (1H, ddd, J=4.6, 8.3, 8.3 Hz), 5.20 (1H, dddd, J=1.0, 6.6, 6.6, 15.4 Hz), 5.32–5.43 (2H, m), 5.54 (1H, dddd, J=1.0, .6.6, 6.6, 15.4 Hz), 5.79 (1H, d, $J=4.6$ Hz), 6.02 (1H, br s), 7.30–7.43(6H, m), 7.63–7.68 (4H, m); ¹³C NMR δ 14.1, 19.3, 20.3, 20.5, 22.6, 22.8, 24.8, 27.1, 28.6, 28.7, 29.2, 31.8, 31.9, 32.2, 32.4, 38.1, 62.6, 62.7, 72.0, 74.7, 81.6, 123.1, 127.2, 127.4, 129.3, 129.4, 130.8, 133.1, 134.7, 135.1, 135.9, 136.0, 168.8, 169.4, 170.1, 172.4; HRMS (FAB) m/z calcd for $C_{43}H_{62}NO_8Si$, $(M+H)^+$ 748.4244, found: 748.4248.

4.5.3. N-{(3S,4R,5R)-4-(Acetyloxy)-5-[(2E,8E,10S)-10-(acetyloxy)- 2,8-hexadecadien-1-yl]-3-[(acetyloxy)methyl]tetrahydro-2-oxo-3 furanyl}-acetamide (28a). By the similar reaction conditions employed for the preparation of 28b from 31, compound 33 (14 mg, 0.018 mmol) was converted into 28a (2.6 mg, 26%): colorless syrup; $[\alpha]_D^{21}$ +30.0 (c 0.22, CHCl₃); IR $\nu_{\rm max}$ (neat) 3350, 2920, 2855, 1790, 1750, 1725, 1680, 1540, 1460, 1375, 1240, 1030 cm⁻¹; ¹H NMR δ 0.87 (3H, t, J=7.0 Hz), 1.22–1.40 (12H, m), 1.50–1.66 (2H, m), 1.99–2.05 (4H, m), 2.02, 2.03, 2.05, and 2.10 (each 3H, 4s), 2.29–2.49 (2H, m), 4.50 and 4.52 (each 1H, 2d, J=11.5 Hz), 4.72 (1H, ddd, J=4.4, 8.0, 8.0 Hz), 5.17 (1H, dt, J=6.8, 6.8 Hz), 5.32-5.44 (2H, m), 5.51-5.71 (2H, m), 5.79 (1H, d, J=4.4 Hz), 5.99 (1H, br s); ¹³C NMR δ 14.0, 20.3, 20.5, 21.4, 22.6, 22.8, 25.2, 28.5, 28.7, 29.0, 31.7, 32.0, 32.2, 32.3, 34.5, 62.6, 62.7, 71.9, 75.0, 81.6, 123.2, 128.6, 134.0, 135.0, 168.8, 169.3, 170.2, 170.4, 172.4; HRMS (FAB) m/z calcd for C₂₉H₄₅NO₉Na, $(M+Na)^+$ 574.2992, found: 574.2988.

4.5.4. 14-epi-Mycestericin A (34). By the similar reaction conditions employed for the preparation of 1 from 28b, compound 28a (4.3 mg, 7.8 μ mol) was converted into 34 (2.5 mg, 81%): amorphous solid; [α] $_{{\rm D}}^{21}$ –5.0 (c 0.15, MeOH); IR $\nu_{\rm max}$ (KBr disk) 3400, 3270, 2920, 2855, 1630, 1460, 1400, 1050, 965 cm $^{-1}$; 1 H NMR (CD₃OD) δ 0.89 $(3H, br t, J=6.8 Hz), 1.25-1.60 (14H, m), 1.93-2.10 (4H, m), 2.26 (2H,$ br t, J=6.8 Hz), 3.77–3.95 (4H, m), 4.00 (1H, d, J=11.0 Hz), 5.34–5.64 (4H, m); ¹³C NMR (CD₃OD) δ 14.4, 23.6, 26.6, 29.95, 30.0, 30.4, 33.0, 33.1, 33.6, 38.5, 38.7, 65.1, 70.5, 71.3, 73.7, 73.8, 126.9, 132.3, 134.59, 134.64 (a signal of the carboxyl carbon could not be detected due to its low intensity); HRMS (FAB) m/z calcd for C₂₁H₄₀NO₆, (M+H)⁺ 402.2856, found: 402.2847.

4.6. Degradation study of mycestericin A and its 14-epimer

4.6.1. (R,E)-Dec-2-ene-1,4-diyl diacetate (35) and its (S,E)-isomer (ent-35). To a mixture of $22b$ (7.2 mg, 0.024 mmol) and MS4A (50 mg) was added TBAF (62 mg, 0.24 mmol) in DMPU (0.4 mL), and the mixture was stirred at 90° C for 3 h. To the mixture at room temperature were added pyridine (1.5 mL) and $Ac₂O$ (1.0 mL) , and the mixture was stirred at room temperature for 19 h. The reaction mixture was diluted with EtOAc, and washed with brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (0.3 g silica gel, 1:15–1:10 EtOAc/hexane as an eluent) to afford **35** (4.7 mg, 77%) as a colorless syrup: $[\alpha]_D^{21}$ +26.5 (c 1.20, CHCl₃); IR ν_{max} (neat) 2960, 2930, 2860, 1740, 1460, 1430, 1370, 1240, 1025 cm⁻¹; ¹H NMR δ 0.87 (3H, t, J=6.6 Hz), 1.24-1.31 (8H, m), 1.49–1.67 (2H, m), 2.05 and 2.06 (each 3H, 2s), 4.54 (2H, d, J=4.9 Hz), 5.24 (1H, dt, J=6.0, 6.8 Hz), 5.67 (1H, dd, J=6.0, 15.6 Hz), 5.77 (1H, td, J=4.9,15.6 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 14.0, 20.9, 21.2, 22.5, 25.0, 29.0, 31.6, 34.2, 64.0, 73.6, 126.3, 132.6, 170.3, 170.6; HRMS (FAB) m/z calcd for C₁₄H₂₅O₄, $(M+H)^+$ 257.1753, found: 257.1751.

By the similar reaction conditions as employed for the synthesis of 35 from 22b, ent-22b (5.0 mg, 0.016 mmol) was converted into ent-**35** (3.2 mg, 75%): [α] $^{21}_{D}$ –26.6 (c 0.23, CHCl₃).

4.6.2. (R)-Octane-1,2-diyl dibenzoate (36) and its (S)-isomer (ent-36). Ozone was introduced into a solution of 35 (13.2 mg, 0.052 mmol) in MeOH (0.8 mL) at -78 °C for 5 min. After confirming the complete consumption of the starting material (TLC analysis), excess ozone was removed with a stream of Ar gas. To the reaction mixture was added NaBH₄ (9.8 mg, 0.26 mmol) at -78 $^{\circ}$ C and the mixture was stirred for 5 min at -78 °C. To this mixture was added K_2CO_3 (36 mg, 0.26 mmol) at -78 °C, and the reaction mixture was stirred at $0 °C$ for 40 min. The reaction mixture was diluted with saturated aqueous NH4Cl solution, and products were extracted with EtOAc. The organic layer was washed with saturated aqueous NH4Cl solution, and dried. Removal of the solvent gave a crude diol. To a solution of the crude diol in pyridine (1.0 mL) at 0 \degree C was added BzCl (46 µL, 0.40 mmol) at 0 \degree C and the mixture was stirred at room temperature for 19 h. The reaction mixture was diluted with saturated aqueous $NH₄Cl$ solution, and products were extracted with EtOAc. The organic layer was washed successively with saturated aqueous NH₄Cl solution, and saturated aqueous $NaHCO₃$ solution, $H₂O$ and brine, and dried. Removal of the solvent left a residue, which was purified by column chromatography (0.3 g silica gel, 1:40 EtOAc/hexane as an eluent) to afford 36 (9.1 mg, 49% for three steps) as a colorless syrup: $\lbrack \alpha \rbrack^{24}_D + 5.4$ (c 0.63, CHCl₃); IR v_{max} (neat) 2960, 2930, 2860, 1740, 1710, 1600, 1460, 1315, 1280, 1240, 1110, 1070, 1025 cm⁻¹; ¹H NMR δ 0.87 (3H, t, J=6.6 Hz), 1.26-1.51 (8H, m), 1.71-1.92 (2H, m), 4.47 (1H, dd, J=6.6, 11.7 Hz), 4.56 $(1H, dd, J=3.4, 11.7 Hz)$, 5.50 $(1H, dd, J=3.4, 6.6, 7.3 Hz)$, 7.38-7.47 (4H, m), 7.51-7.58 (2H, m), 7.99-8.07 (4H, m); ¹³C NMR δ 14.0, 22.5, 25.2, 29.1, 31.0, 31.6, 65.7, 72.3, 128.4, 129.7, 132.95, 133.01, 166.1; Retention time of HPLC [DAICEL CHIRLCEL OJ-H, 4.6 mm ID \times 250 mm, *i*-PrOH/hexane=1:300, flow rate=0.8 mL/min, UV (254 nm) detection], 21.9 min ($>99\%$ ee); HRMS (FAB) m/z calcd for $C_{22}H_{27}O_4$, $(M+H)^+$ 355.1909, found: 355.1904.

By the similar reaction conditions as employed for the synthesis of 36 from 35, ent-35 (8.5 mg, 0.033 mmol) was converted into ent-**36** (5.3 mg, 45%): $[\alpha]_D^{21}$ –6.0 (c 0.15, CHCl₃); Retention time of HPLC [DAICEL CHIRLCEL OI-H, 4.6 mm ID \times 250 mm, *i*-PrOH hexane=1:300, flow rate=0.8 mL/min, UV (254 nm) detection], 19.0 min (>99% ee).

4.6.3. Degradation of synthetic mycestericin $A(1)$. To a solution of mycestericin A (1) $(2.5 \text{ mg}, 6.2 \text{ µmol})$ in pyridine (1.5 mL) was added Ac_2O (1.0 mL), and the mixture was stirred at room temperature for 15 h. The reaction mixture was concentrated to give a residue, which was purified by column chromatography (0.3 g silica gel, 1:2–1:1 EtOAc/hexane as an eluent) to afford tetraacetyl mycestericin A γ -lactone (28b) (2.5 mg, 73%) as a colorless syrup. Ozone was introduced into a solution of $28b$ (2.5 mg, 4.5 µmol) in MeOH (1.5 mL) at -78 °C for 5 min. After confirming the complete consumption of the starting material (TLC analysis), excess ozone was removed with a stream of Ar gas. To the reaction mixture was added NaBH₄ (0.5 mg 13.2 µmol) at -78 °C. After being stirred for 5 min, the reaction was quenched by addition of 1 M aqueous HCl solution (0.1 mL) and diluted with water. Products were extracted with EtOAc. The organic layer was washed with 1 M aqueous HCl solution, and dried. Removal of the solvent left a residue, which was roughly purified by column chromatography (0.1 g silica gel, 1:7 EtOAc/hexane as an eluent). The active fractions were collected and concentrated to give an oil, which was dissolved in MeOH (1.0 mL). To this solution at $0 °C$ was added K_2CO_3 (1.8 mg, 13.2 μ mol). After being stirred for 30 min, the reaction mixture was diluted with saturated aqueous NH4Cl solution, and products were extracted with EtOAc. The organic layer was washed with saturated NH4Cl aqueous, and dried. Removal of the solvent gave crude octane-1,2-diol. To a solution of crude octane-1,2-diol in pyridine (1.0 mL) was added BzCl (50 μ L, 380 μ mol) at 0 °C. The mixture was stirred at room temperature for 13 h. The reaction

mixture was diluted with saturated NH4Cl aqueous, and products were extracted with EtOAc. The organic layer was washed successively with saturated aqueous NH4Cl solution, and saturated aqueous NaHCO₃ solution, H₂O and brine, and dried. Removal of the solvent left a residue, which was purified by preparative TLC (1:50 EtOAc/hexane, developed three times) to afford 36 (1.1 mg, 50% for four steps) as a colorless syrup: HRMS (FAB) m/z calcd for $C_{22}H_{27}O_4$, $(M+H)^+$ 355.1909, found: 355.1908. The ¹H NMR spectral data were fully identical with those of 36 prepared from 35, and chiral HPLC analysis assigned the absolute configuration of 36 prepared from 1 as R with $>99\%$ ee.

4.6.4. Degradation of 14-epi-mycestericin A (34). The similar reaction conditions as described for conversion of 1 to 36 were applied to 14-epi-mycestericin A $(34, 1.5 \text{ mg}, 3.7 \text{ mmol})$ to afford ent-36 (0.5 mg, 38% for four steps) as a colorless syrup: HRMS (FAB) m/z calcd for C₂₂H₂₇O₄, (M+H)⁺ 355.1909, found: 355.1907. The ¹H NMR spectral data were fully identical with those of 36 prepared from 35, and chiral HPLC analysis assigned the absolute configuration of ent-**36** prepared from **34** as S with $>99\%$ ee.

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References and notes

- 1. (a) Sasaki, S.; Hashimoto, R.; Kiuchi, M.; Inoue, K.; Ikumoto, T.; Hirose, R.; Chiba, K.; Hoshino, Y.; Okumoto, T.; Fujita, T. J. Antibiot. 1994, 47, 420-433; (b) Fujita, T.; Hamamichi, N.; Kiuchi, M.; Matsuzaki, T.; Kitao, Y.; Inoue, K.; Hirose, R.; Yoneta, M.; Sasaki, S.; Chiba, K. J. Antibiot. 1996, 49, 846–853.
- 2. (a) Klurpfel, D.; Bagli, J.; Baker, H.; Charest, M.-P.; Kudelski, A.; Sehgal, S. N.; Vézina, C. J. Antibiot. 1972, 25, 109-115; Bagli, J. F.; Kluepfel, D.; Jacques, M. S. J.Org. Chem. 1973, 38, 1253–1260; (b) Aragozzini, F.; Manachini, P. L.; Craveri, R.; Rindone, B.; Scolastico, C. Tetrahedron 1972, 28, 5493–5498; (c) Fujita, T.; Inoue, K.; Yamamoto, S.; Sasaki, S.; Toyama, R.; Yoneta, M.; Hoshino, Y.; Okumoto, T. J. Antibiot. 1994, 47, 208–215; Fujita, T.; Inoue, K.; Yamamoto, S.; Ikumoto, T.; Sasaki, S.; Toyama, R.; Yoneta, M.; Chiba, K.; Hoshino, Y.; Okumoto, T. J. Antibiot. 1994, 47, 216–225; Sasaki, S.; Hashimoto, R.; Kiuchi, M.; Inoue, K.; Ikumoto, T.; Hirose, R.; Chiba, K.; Hoshino, Y.; Okumoto, T.; Fujita, T. J. Antibiot. 1994, 47, 420–433.
- 3. Brunner, M.; Koskinen, A. M. P. Curr. Org. Chem. 2004, 8, 1629–1645.
- 4. For isolation of sphingofungins A–D, see: (a) VanMiddlesworth, F.; Giacobbe, R. A.; Lopez, M.; Garrity, G.; Bland, J. A.; Batizal, K.; Frontling, R. A.; Wilson, K. E.; Monaghan, R. L. J. Antibiot. 1992, 45, 861–867; (b) VanMiddlesworth, F.; Dufresne, C.; Wincott, F. E.; Mosley, R. T.; Wilson, K. E. Tetrahedron Lett. 1992, 33, 297–300 For isolation of sphingofungins E and F, see: (c) Horn, W. S.; Smith, J. L.; Bills, G. F.; Raghoobar, S. L.; Helms, G. L.; Kurta, M. B.; Marrinan, J. A.; Frommer, B. R.; Thornton, R. A.; Mandala, S. M. J. Antibiot. 1992, 45, 1692– 1696.
- 5. Yamaji-Hasegawa, A.; Takahashi, A.; Tetsuka, Y.; Senoh, Y.; Kobayashi, T. Biochemistry 2005, 44, 268–277.
- 6. (a) Miyake, Y.; Kozutumi, Y.; Nakamura, S.; Fujita, T.; Kawasaki, T. Biochem. Biophys. Res. Commun. 1995, 211, 396–403; (b) Zweerink, M. M.; Edison, A. M.; Wells, G. B.; Pinto, W.; Lester, R. L. J. Biol. Chem. 1992, 267, 25032–25038.
- 7. Recent reviews of synthesis of natural products possessing α -substituted a-amino acid structures, see: (a) Kang, S. H.; Kang, S. Y.; Lee, H.-S.; Buglass, A. J. Chem. Rev. 2005, 105, 4537–4558; (b) Ohfune, Y.; Shinada, T. Eur. J. Org. Chem. 2005, 5127-5143; (c) Cativiela, C.; Díaz-de-Villegas, M. D. Tetrahedron: Asymmetry 2007, 18, 569–623.
- 8. Recent reviews on chemistry and biological activities of myriocin-related compounds, see: (a) Byun, H.-S.; Lu, X.; Bittman, R. Synthesis 2006, 2447–2474; (b) Liao, J.; Tao, J.; Lin, G.; Liu, D. Tetrahedron 2005, 61, 4715–4733.
- 9. For total synthesis of myriocin, see: (a) Banfi, L.; Beretta, M. G.; Colombo, L.; Gennari, C.; Scolastico, C. J. Chem. Soc., Chem. Commun. 1982, 488–490; J. Chem. Soc., Perkin Trans. 1 1983, 1613–1619; (b) Yoshikawa, M.; Yokokawa, Y.; Okuno, Y.; Murakami, N. Chem. Pharm. Bull. 1994, 42, 994-996; Tetrahedron 1995, 51, 6209–6228; (c) Sano, S.; Kobayashi, Y.; Kondo, T.; Takebayashi, M.; Maruyama, S.; Fujita, T.; Nagao, Y. Tetrahedron Lett. 1995, 36, 2097–2100; (d) Hatakeyama, S.; Yoshida, M.; Esumi, T.; Iwabuchi, Y.; Irie, H.; Kawamoto, T.; Yamada, H.; Nishizawa, M. Tetrahedron Lett. 1997, 38, 7887–7890; (e) Oishi, T.; Ando, K.; Chida, N. Chem. Commun. 2001, 1932–1933; (f) Lee, K.-Y.; Oh, C.-Y.; Kim, Y.-H.; Joo, J.-E.; Ham, W.-H. Tetrahedron Lett. 2002, 43, 9361–9363; (g) Martinkova´, M.; Gonda, J.; Raschmanová, J.; Vojticková, M. Tetrahedron 2007, 63, 10603– 10607; (h) Jones, M. C.; Marsden, S. P. Org. Lett. 2008, 10, 4125–4128; (i) Inai, M.;

Toto, T.; Furuta, T.; Wakimoto, T.; Kan, T. Tetrahedron: Asymmetry 2008, 19, 2771–2773.

- 10. For total synthesis of sphingofungins E and/or F, see: (a) Kobayashi, S.; Matsumura, M.; Furuta, T.; Hayashi, T.; Iwamoto, S. Synlett 1997, 301-303; (b) Kobayashi, S.; Furuta, T.; Hayashi, T.; Nishijima, M.; Hanada, K. J. Am. Chem. Soc. 1998, 120, 908–919; (c) Kobayashi, S.; Furuta, T. Tetrahedron 1998, 54, 10275–10294; (d) Trost, B. M.; Lee, C. B. J. Am. Chem. Soc. 1998, 120, 6818–6819; Trost, B. M.; Lee, C. J. Am. Chem. Soc. 2001, 123, 12191–12201; (e) Liu, D.-G.; Wang, B.; Lin, G.-Q. J. Org. Chem. 2000, 65, 9114–9119; (f) Wang, B.; Yu, X.-M.; Lin, G.-Q. Synlett 2001, 904–906; (g) Nakamura, T.; Shiozaki, M. Tetrahedron Lett. 2001, 42, 2701–2704; Tetrahedron 2002, 58, 8779–8791; (h) Oishi, T.; Ando, K.; Inomiya, K.; Sato, H.; Iida, M.; Chida, N. Org. Lett. 2002, 4, 151–154; Bull. Chem. Soc. Jpn. 2002, 75, 1927–1947; (i) Lee, K.-Y.; Oh, C.-Y.; Ham, W.-H. Org. Lett. 2002, 4, 4403–4405; (j) Li, M.; Wu, A. Synlett 2006, 2985–2988.
- 11. For total synthesis of mycestericins, see: (a) Fujita, T.; Hamamichi, N.; Matsuzaki, T.; Kitao, Y.; Kiuchi, M.; Node, M.; Hirose, R. Tetrahedron Lett. 1995, 36, 8599–8602; (b) Shibata, K.; Shingu, K.; Vassilev, V. P.; Nishide, K.; Fujita, T.; Node, M.; Kajimoto, T.; Wong, C.-H. Tetrahedron Lett. 1996, 37, 2791–2794; (c) Nishide, K.; Shibata, K.; Fujita, T.; Kajimoto, T.; Wong, C.-H.; Node, M.
Heterocycles **2000**, 52, 1191–1201; (d) Iwabuchi, Y.; Furukawa, M.; Esumi, T.; Hatakeyama, S. Chem. Commun. 2001, 2030–2031. Synthesis and biological activity of sulfamisterin and its derivatives, see: (e) Sato, H.; Maeba, T.; Yanase, R.; Yamaji-Hasegawa, A.; Kobayashi, T.; Chida, N. J. Antibiot. 2005, 58, 37–49.
- 12. A part of this work has been published as a preliminary communication, see: Sato, H.; Sato, K.; Iida, M.; Yamanaka, H.; Oishi, T.; Chida, N. Tetrahedron Lett. 2008, 49, 1943–1947.
- 13. Chida, N.; Takeoka, J.; Tsutsumi, N.; Ogawa, S. J. Chem. Soc., Chem. Commun.
1995, 793–794; Chida, N.; Takeoka, J.; Ando, K.; Tsutsumi, N.; Ogawa, S. Tetrahedron 1997, 53, 16287–16298.
- 14. For a comprehensive review on Overman rearrangement and related reactions, see: Overman, L. E.; Carpenter, N. E. In Organic Reactions; Overman, L. E., Ed.; Wiley: New York, NY, 2005; Vol. 66, pp 1–107.
- 15. Recent reports on syntheses of natural products and related compounds utilizing Overman rearrangement, see: (a) Momose, T.; Kaiya, Y.; Hasegawa, J.; Sato, T.; Chida, N. Synthesis 2009, 2983–2991; (b) Imaoka, T.; Iwamoto, O.; Noguchi, K.; Nagasawa, K. Angew. Chem., Int. Ed. 2009, 48, 3799–3801; (c) Hama, N.; Matsuda, T.; Sato, T.; Chida, N. Org. Lett. 2009, 11, 2687–2690; (d) Dickson, D. P.; Wardrop, D. J. Org. Lett. 2009, 11, 1341–1344; (e) Momose, T.; Hama, N.; Higashino, C.; Sato, H.; Chida, N. Tetrahedron Lett. 2008, 49, 1376–1379; (f) Matveenko, M.; Kokas, O. J.; Banwell, M. G.; Willis, A. C. Org. Lett. 2007, 9, 3683–3685; (g) Hakansson, A. E.; Palmelund, A.; Holm, H.; Madsen, R. Chem.-Eur. J. 2006, 12, 3243-3253; (h) Tsujimoto, T.; Nishikawa, T.; Urabe, D.; Isobe, M. Synlett 2005, 433–436; (i) Nishikawa, T.; Urabe, D.; Yoshida, K.; Iwabuchi, T.; Asai, M.; Isobe, M. Chem.-Eur. J. 2004, 10, 452-462; (j) Ohyabu, N.; Nishikawa, T.; Isobe, M. J. Am. Chem. Soc. 2003, 125, 8798–8805.
- 16. Sánchez-Sancho, F.; Valverde, S.; Herradón, B. Tetrahedron: Asymmetry 1996, 7, 3209–3246.
- 17. Nishikawa, T.; Asai, M.; Ohyabu, N.; Isobe, M. J. Org. Chem. 1998, 63, 188–192.
- 18. The optical purities of 14 , $5a$, and $5b$ were all determined to be >99% ee, by chiral column analyses (Chiralcel OD-H), confirming that no racemization had occurred during the preparation of these compounds. See [Experimental](#page-5-0) section.
- 19. Keck, G. E.; Boden, E. P. Tetrahedron Lett. 1984, 25, 265–268; Marshall, J. A. Chem. Rev. 2000, 100, 3163–3186.
- 20. Ohba, S.; Sato, H.; Iida, M.; Chida, N. Acta Crystallogr., Sect. E 2003, E59, o1259o1260.
- 21. (a) Takai, K.; Nitta, K.; Utimoto, K. J. Am. Chem. Soc. 1986, 108, 7408–7410; (b) Evans, D. A.; Black, W. C. J. Am. Chem. Soc. 1993, 115, 4497–4513.
- 22. (a) Carmack, M.; Kelly, C. J. J. Org. Chem. 1968, 33, 2171–2173; (b) Rubin, L. J.; Lardy, H. A.; Fischer, H. O. L. J. Am. Chem. Soc. 1952, 74, 425–428.
- 23. Narisada, M.; Ohtami, M.; Watanabe, F.; Uchida, K.; Arita, H.; Doteuchi, M.; Hanasaki, K.; Kakushi, H.; Otani, K.; Hara, S. J. Med. Chem. 1988, 31, 1847–1854. 24. Taber, D. F.; Deker, P. B.; Gaul, M. D. J. Am. Chem. Soc. 1987, 109, 7488–7494.
- 25. (a) Negishi, E. Pure Appl. Chem. 1981, 53, 2333-2356; (b) Negishi, E. Acc. Chem. Res. 1982, 15, 340–348; (c) Williams, D. R.; Kissel, W. S. J. Am. Chem. Soc. 1998, 120, 11198–11199; (d) Tamaru, Y.; Ochiai, H.; Nakamura, T.; Yoshida, Z.Angew. Chem., Int. Ed. Engl.1987, 26, 1157–1158; (e) Negishi, E. Bull. Chem. Soc. Jpn. 2007, 80, 233–257.
- 26. The structure of 25 has been tentatively assigned; there is a possibility that compound 25 would be a positional isomer of the conjugated carbon–carbon double bonds.
- 27. Isobe, M.; Ichikawa, Y.; Bai, D.-L.; Masaki, H.; Goto, T. Tetrahedron 1987, 43, 4767–4776.
- 28. Lipshutz, B. H.; Miller, T. A. Tetrahedron Lett. 1989, 30, 7149–7152 Significant amounts of the corresponding (ethoxymethyl)ether were formed when 31 was reacted with Bu4NF in THF or CsF in HMPA.
- 29. The low yield of this step was mainly due to the decomposition of the substrate; the formation of polar by-products, whose structures could not be determined, was observed.
- 30. For a review on the Suzuki–Miyaura coupling, see: (a) Miyaura, N.; Suzuki, A. Chem. Rev. 1995, 95, 2457–2483 For a review on the B-alkyl Suzuki–Miyaura coupling, see: (b) Chemler, S. R.; Trauner, D.; Danishefsky, S. J. Angew. Chem., Int. Ed. 2001, 40, 4544–4568.
- 31. Johnson, C. R.; Braun, M. P. J. Am. Chem. Soc. 1993, 115, 11014–11015.
- 32. The optical purities of 23 c and ent-23 c were determined to be >99% ee, by chiral column analyses (Chiralcel OD-H) of the derived alcohols 24c and ent-24c. See [Experimental](#page-5-0) section.